

RECOMMENDATIONS

1. Protection is better than treatment so cereals storage must be under healthy conditions of temperature, moisture, away from insects and birds, and not stored for a long period of time than few months in order to avoid formation and spread of fungal secondary metabolites.
2. Buy seeds from Known, reliable, sources where you know it is fresh clean and has been handled properly.
3. It is not recommended to add maize to wheat in Egyptian bread production since it may increase the daily intake of aflatoxin B₁.
4. The use of gamma irradiation is recommended in cereals since it is a safe technology to ensure food security and safety and to overcome strict quarantine quality standard in international trade.
5. Further researches on the methods of reduction or removal of different types of mycotoxins is recommended.
6. Health authorities should make regular inspections to detect the amount of aflatoxin in food.
7. Strict laws should be applied to prohibit the use of aflatoxins contaminated materials in food industries.

SUMMARY

Occurrence of mycotoxins in cereal foodstuff is unavoidable. The attendant challenges to crop production, yield and quality loss is about 25% annually according to FAO.

Aflatoxin B₁ is the most potent hepatocarcinogen known in animals and it is classified by the International Agency of Research on Cancer (IARC) as Group I carcinogen meaning that it is a proven cancer-inducing agent. It also occurs in the environment contaminating a lot of different food and feed commodities. Aflatoxins are a group of mycotoxins produced in tropical and sub-tropical regions. Aflatoxins are hepatic and carcinogenic secondary metabolites of moulds that produce mainly from *Aspergillus flavus* and *Aspergillus paraciticus*. Aflatoxins contaminate a variety of agricultural commodities in countries with hot and humid climates.

The aim of this study was to assess the possible effect of gamma irradiation on the reduction of aflatoxin B₁ in some cereal grains and the impact on nutritive values including, ash, moisture, fat, protein, carbohydrates, crude fibers, calories, fatty acids profiles and amino acid profiles.

One group pre and post intervention study was performed on 60 samples (one kilogram for each sample) divided equally among the three cereals including maize, wheat, and rice.

To achieve this aim the following was carried out:

Samples (maize, wheat, and rice) were randomly selected from the local markets and wholesale markets in Alexandria city during June, July, and August-2013. Each one kilogram sample was subdivided into equal four sub-groups. Samples were stored in plastic bags at 4° C until analysis.

- A.** The first quarter quantity of each cereal sub-sample (control sample) was taken to the central lab of High Institute of Public Health for the following analysis:-
1. Determination of aflatoxin B₁ by high performance liquid chromatography.
 2. Proximate analysis for cereal samples include:
 - Determination of moisture contents
 - Determination of fat contents by Soxhlet method.
 - Determination of protein contents by Kjeldahl nitrogen method.
 - Determination of ash contents.
 - Calculation of carbohydrate contents.
 - Determination of crude fibers
 - Calculation of sample calories.
 3. Preparation of methyl esters by sodium methoxide method followed by determination of fatty acid profile by gas liquid chromatography.
 4. Determination of amino acid profile by amino acid analyzer.

-
- B.** The second quarter sub-sample of each cereal type was irradiated using gamma irradiation source from ^{60}CO at an absorbed dose level of 4 kGy each followed by analysis for aflatoxin B₁, proximate analysis, fatty acid profile and amino acid profile, analysis was performed as described previously.
- C.** The third quarter sub-sample of each cereal type was irradiated using gamma irradiation from ^{60}CO at an absorbed dose level of 6 kGy each followed by analysis for aflatoxin B₁, proximate analysis, fatty acid profile and amino acid profile as described previously.
- D.** The fourth quarter sub-sample of each cereal type was irradiated using gamma irradiation from ^{60}CO at an absorbed dose level of 8 kGy each followed by analysis for aflatoxin B₁, proximate analysis, fatty acid profile and amino acid profile as described previously.

The results of this study was statistically analyzed; data were fed to the computer and analyzed using IBM SPSS software package version 20.0 and significant data was taken at $p < 0.05$ using ANOVA test.

It was found that:

1. Maize samples contain the highest level of aflatoxin B₁ than wheat and rice.
2. Gamma irradiation is a suitable technique which reduces the levels of aflatoxin B₁ in cereal samples without affecting the nutritive values, at **4 KGy** the reduction percents of aflatoxin B₁ were 15.54%, 22.25%, and 27.46% for maize, wheat, and rice, respectively whereas at **6 KGy** the reduction percents of aflatoxin B₁ were 32.39%, 43.84%, and 56.38% for maize, wheat, and rice, respectively and the **8 KGy** radiation dose removed about 60.26% of the toxin in maize, 64.68% in rice and 69.29% in wheat samples.
3. Higher radiation doses than 8 KGy are required to remove the toxin until it reaches the legal limit (5ppb) according to FAO.
4. By increasing the radiation doses the percents of reduction of aflatoxin B₁ increase (dose dependent manner) in cereal samples.
5. The percents of reduction decrease with increasing the oil contents of cereals so maize had the lowest aflatoxin B₁ reduction level.
6. Gamma irradiation affect on fatty acid profiles of wheat and rice, causes reduction in mono-unsaturated fatty acid (oleic acid, C18:1) and poly-unsaturated fatty acid (linolenic acid, C18:3) which disappear completely in rice at 8 KGy, while in maize samples there was increase in the saturated fatty acid (palmitic acid, C16:0) and decrease in the mono-unsaturated fatty acid (oleic acid, C18:1) and this consistent with other studies.
7. Gamma irradiation affects slightly on amino acid profile of cereal grains in **maize** samples there is a reduction in phenyl alanine and threomine and increase in leucine, in **wheat** samples there is a decrease in tryptophan, isoleucine, phenylalanine, arginine and lysine and an increase in leucine and in **rice** samples there is increase in leucine and decrease in phenylalanine.

REFERENCES

1. Wild CP, Hall AJ. Primary prevention of hepatocellular carcinoma in developing countries. *Mutat Res* 2000; 462(2-3): 381-93.
2. Anwar WA, Khaled HM, Amra HA, El-Nezami H, Loffredo CA. Changing pattern of hepatocellular carcinoma (HCC) and its risk factors in Egypt: Possibilities for prevention. *Mutat Res* 2008; 659: 176–84.
3. Paterson RRM, Lima N. Further mycotoxin effects from climate change. *Food Res Int* 2010; 43: 1902-14.
4. Kozakiewicz Z, Smith D. Physiology of *Aspergillus*. In: *Aspergillus*. Smith JE (ed). Plenum Press, New York. 1994, p.23.
5. Chapman GP. The biology of grasses. Centre for Agricultural Bioscience International, Wallingford, Oxon, UK. 1996, p.273.
6. Evans EJ. Cereal production methods. In: *Cereals processing technology*. Owen G (ed). Woodhead Publishing Limited Abington Hall, Abington Cambridge England. 2001, pp.23-5.
7. Gwary OM, Hati SS, Dimari GA, Ameh JA. Assessment of mycotoxins (Total aflatoxins and ochratoxin-A) contamination of staple cereals. *IJCBS* 2012; 2: 1-6.
8. Council for Agricultural Science and Technology (CAST). *Mycotoxins: economic and health risks*, task force sheet No 116. Ames, Iowa: CAST; 1989.
9. D’Mello JPF, MacDonald AMC. Mycotoxins. *Anim Feed Sci Tech* 1997; 69: 155–66.
10. World Health Organization (WHO), International Agency of Research on Cancer (IARC). *IARC monographs on the evaluation of carcinogenic risks to humans some traditional herbal medicines, some mycotoxins, naphthalene and styrene*. Volume 82. Lyon: IARC Working Group on the Evaluation of Carcinogenic Risks to Humans; 2002.
11. Blount WP. Turkey X disease. *Turkeys* 1961; 9: 55-8.
12. Bullerman BL, Bianchini A. Stability of mycotoxins during food processing. *Int J Food Microbiol* 2007; 119(1-2): 140-6.
13. Riazipour M, Fooladi AAI, Bagherpour G. Survey of T-2 toxin present in cereals destined for human consumption. *Jundishapur J Microbiol* 2012; 5(3): 497-501.
14. L-park D. Effect of processing on aflatoxin. In: *Mycotoxins and food safety*. Devries JW, Trucksess MW, Jackson L (eds). Kluwer Academic, Plenum Publishers. 2002, pp.173-9.

15. Wagacha JM, Muthomi JW. Mycotoxin problem in Africa: Current status, implications to food safety and health and possible management strategies. *Int J Food Microbiol* 2008; 124(1): 1–12.
16. Fandohan P, Zoumenou D, Hounhouigan DJ, Marasas WF, Wingfield MJ, Hell K. Fate of aflatoxins and fumonisins during the processing of maize into food products in Benin. *Int J Food Microbiol* 2005; 98(3): 249-59.
17. Tian J, Ban X, Zeng H, He J, Huang B, Wang Y. Chemical composition and antifungal activity of essential oil from *Cicuta virosa* L. var. *latisepta* Celak. *Int J Food Microbiol* 2011; 145(2): 464-7.
18. Van Dyck PJ, Tobback P, Feyes M, Van de Voorde H. Sensitivity of aflatoxin B1 to ionizing radiation. *Appl Environ Microbiol* 1982; 43(6): 1317-9.
19. Fapohunda SO, Anjorin ST, Adesanmi CA. Nutritional profile of gamma-irradiated and non-irradiated *Sesamum indicum* seeds from Abuja markets. *J Anim Prod Adv* 2012; 2(3): 161-5.
20. Hesseltine CW. Conditions leading to mycotoxin contamination of foods and feeds. In: *Mycotoxins, other fungal related food problems*. Rodricks JV (ed). American Chemical Society, Washington DC 1976, pp.1-22.
21. Frisvad JC, Smedsgaard J, Larsen TO, Samson RA. Mycotoxins, drugs and other extrolites produced by species in *Penicillium* subgenus *Penicillium*. *Stud Mycol* 2004; 49: 201–42.
22. Masoero F, Gallo A, Moschini M, Piva G, Díaz D. Carryover of aflatoxin from feed to milk in dairy cows with low or high somatic cell counts. *Animal* 2007; 1:1344–50.
23. Strosnider H, Azziz-Baumgartner E, Banziger M, Bhat RV, Breiman R. Public health strategies for reducing aflatoxin exposure in developing countries: A work group report. *Environ Health Persp* 2006; 12:1898-903.
24. Forgacs J and Carli WT. Mycotoxicoses. *Adv Vet Sci* 1962; 7:273.
25. Sargeant K, O'Kelly J, Carnaghan RBA, Alicroft R. The assay of a toxic principle in certain groundnut meals. *Vet. Rec* 1961; 73: 1219-23.
26. Sargeant K, Sheridan A, O'Kelly J, Carnaghan RBA. Toxicity associated with certain samples of groundnuts. *Nature* 1961; 192: 1096-7.
27. Lancaster MC, Jenkins FP Philp J. Toxicity associated with certain samples of groundnuts. *Nature* 1961; 192: 1095-96.
28. Fink-Gremmels J. Mycotoxins: Their implications for human and animal health. *Vet. Quart* 1999; 21: 115-20.

29. Kuiper-Goodman T. Food safety: Mycotoxins and phycotoxins in perspective. In: Mycotoxins and phycotoxins developments in chemistry, toxicology and food safety. Miraglia M, Van Egmond HP, Brera C, Gilbert J (eds). Alaken Inc., Fort Collins, Colorado. 1998, pp. 25-48.
30. Bennett JW, Klich M. Mycotoxins. Clin. Microbiol 2003; 16:497-516.
31. Hormisch D, Brost I, Kohring GW, Giffhorn F, Kroppenstedt RM, Stackebrandt E, et al. *Myobacterium fluoranthenivorans* sp. nov., a fluoranthene and aflatoxin B₁ degrading bacterium from contaminated soil of a former coal gas plant. Syst. Appl. Microbiol 2004; 27: 653- 60.
32. Aquino S, Goncalvez E, Ries TA, Sabundjian IT, Trindade RA, Rossi MH, et al. Effect of γ -irradiation on mycoflora of guarana (*Paullinia cupana*). Rad Phys Chem 2007; 76: 1470-3.
33. Boffetta P. Biomarkers in cancer epidemiology: an integrative approach. Carcinogenesis 2010; 31(1): 121-6.
34. Wogan GN. Chemical nature and biological effects of the aflatoxins. Bacteriol Rev 1966; 30(2): 460.
35. Yabe K, Matsushima K, Koyama T, Hamasaki T. Purification and characterization of o-methyltransferase involved in conversion of demethylsterigmatocystin to sterigmatocystin and of dihydrodemethyl-sterigmatocystin to dihydrosterigmatocystin during aflatoxin biosynthesis. Appl Envir Microbiol 1998; 64(1): 166–71.
36. Jideani IA. Traditional and possible technological uses of *Digitaria exilis* (acha) and *Digitaria iburu* (iburu): A review. Plant Foods Hum Nutr 1999; 54: 363-74.
37. Ihekoronye AI, Ngoddy PO. Integrated food chemical analysis of Food. 8th Ed. London: Churchill Living Stone. 1985, pp.446-69.
38. Anthony MH, Francis DM, Berka NP, Ayinla GT, Haruna OG. In: Trends in vital food and control engineering. Eissa AA (ed). In Tech. DOI. 2012, PP.196-9.
39. Kent WL. Technology of cereal. Pergamon (ed), Headington Hill Hall, Oxford UK. 1983, pp.49-60.
40. Atehnkeng J, Ojiambo PS, Donner M, Ikotun K, Sikora RA, Cotty PJ, Bandyopadhyay R. Distribution and toxicity of *Aspergillus* species isolated from maize kernels from three agroecological zones of Nigeria. Int J Food Microbiol 2008; 122: 74–84.
41. Essono G, Ayodele M, Akoa A, Foko J, Filtenborg O, Olembo S. Aflatoxin producing *Aspergillus* spp. and aflatoxin levels in stored cassava chips as affected by processing practice. Food Control 2009; 20: 648–54.
42. Njobeh BP, Dutton MF, Koch SH, Chuturgoon A. Contamination with storage fungi of human foods from Cameroon. Int J Food Microbiol 2009; 135: 193-8.

43. Makun HA, Dutton MF, Njobeh PB, Mwanza M, Kabiru AY. Natural multi-mycotoxin occurrence in rice from Niger State. *Nigeria Mycotoxin Research* 2011; 27 (2): 97-104.
44. Food and Agriculture Organization of the United Nations (FAO). Post harvest losses in quality of food grains. Food and Nutrition Paper No 29. Rome, Italy: FAO; 1983.
45. Stössel P. Aflatoxin contamination in soybeans: Role of proteinase inhibitors, zinc availability, and seed coat integrity. *Appl Environ Microbiol* 1986; 52: 68–72.
46. Schindler AF. Temperature limits for production of aflatoxin by twenty-five isolates of *Aspergillus flavus* and *Aspergillus parasiticus*. *J Food Protect* 1977; 40: 39–40.
47. Ominski KH, Marquardt RR, Sinha RN, Abramson D. Ecological aspects of growth and mycotoxin production by storage fungi. In: *Mycotoxins in grains: Compounds other than aflatoxins*. Miller JD, Trenholm HL (eds). Eagan Press, St. Paul Minnesota, USA. 1994, pp.287-314.
48. Codex Alimentarius Commission. Code of practice for the prevention and reduction of aflatoxin contamination in peanuts. Retrieved 2011.
49. Hell K, Cardwell KF, Setamou M, Poehling HM. The influence of storage practices on aflatoxin contamination in maize in four agroecological zones of Benin West Africa. *J Stored Prod Res* 2000; 36: 365–82.
50. Bilgrami KS, Prasad T, Misra RS, Sinha KK. Aflatoxin contamination in maize under field conditions. *Indian Phytopath* 1981; 34: 67–8.
51. Denning DW, Riniotis K, Dobrashiart R, Sambatakou H. Chronic cavitary and fibrosing pulmonary and pleural aspergillosis: Case series, proposed nomenclature and review. *Clin Infect Dis* 2003; 37 (3): 265-80.
52. Ronning CM, Fedorova ND, Bowyer P. Genomics of *Aspergillus fumigatus*. *Rev Iberoam Micol* 2005; 22(4): 223-8.
53. Stevens DA. Diagnosis of fungal infections current status. *J Antimicrob Chemother* 2002; 49: 11-9.
54. Hocking AD, Doyle MP, Beuchat MR .Toxigenic *Aspergillus* species. In: *Food microbiology fundamentals and frontiers*. Montville TJ (ed). ASM Press, Washington, DC. 1997.
55. Wogan GN. Aflatoxins as risk factors for hepatocellular carcinoma in humans: Aflatoxin exposure and its relationship to kwashiorkor in African children. *Cancer Res* 1992; 52: 2114-8.
56. Lancaster MD, Jenkins FP, Phillip JM. Toxicity associated with certain samples of groundnuts. *Nature* 1961; 192: 1095-6.
57. Center for Disease Control and Prevention (CDC). Outbreak of aflatoxin poisoning--eastern and central provinces, Kenya, January-July 2004. *MMWR Morb Mortal Wkly Rep* 2004; 53(34): 790-3.

58. Fung F, Clark RF. Health effects of mycotoxins: A toxicological overview. *J Toxicol. Clin. Toxicol* 2004; 42: 217-34.
59. Lewis L, Onsongo M, Njapau H, Kenya Aflatoxicosis Investigation Group. Aflatoxin contamination of commercial maize products during an outbreak of acute aflatoxicosis in Eastern and central Kenya. *Res* 2005; 113: 1763-7.
60. Williams JH, Phillips TD, Jolly PE, Stiles JK, Jolly CM, Aggarwal D. Human aflatoxicosis in developing countries: A review of toxicology, exposure, potential health consequences, and interventions. *Am J Clin Nutr* 2004; 80: 1106–22.
61. Robens JF, Richard JL. Aflatoxins in animal and human health. *Rev Environ Contam Toxicol* 1992; 127: 69-94.
62. Bejarano RRJ, Centeno BSJ. Extracto de Citrus limon para el control de aflatoxinas y hongos aflatoxigénicos en alimentos concentrados para pollos de engorde producidos en Venezuela. *Revista de la Sociedad Venezolana de Microbiología* 2009; 29(1): 57-61.
63. Dorner JW, Cole RJ, Diener UL. The relationship of *Aspergillus flavus* and *Aspergillus parasiticus* with reference to production of aflatoxins and cyclopiazonic acid. *Mycopathologia* 1984; 87(1-2): 13-15.
64. Sánchez E, Heredia N, García S. Inhibition of growth and mycotoxin production of *Aspergillus flavus* and *Aspergillus parasiticus* by extracts of A gave species. *Int J Food Microbiol* 2005; 98(3): 271-9.
65. Bressac B, Kew M, Wands J, Ozturk M. Selective G to T mutations of p53 gene in hepatocellular carcinoma from southern Africa. *Nature* 1991; 350: 429-31.
66. Hsu IC, Metcalf RA, Sun T, Welsh JA, Wang NJ, Harris CC. Mutational hot spot in the p53 gene in human hepatocellular carcinomas. *Nature* 1991; 350: 427-8.
67. Moss MO. Risk assessment for aflatoxins in foodstuffs. *Int Biodeter Biodegr* 2002; 50(3-4): 137-42.
68. Kelly JD, Eaton DL, Guengerich FP, Coulombe RJ. Aflatoxin B₁ activation in human lung. *Toxicol Appl Pharmacol* 1997; 144: 88 –95.
69. Dragan YP, Pitot HC. Aflatoxin carcinogenesis in the context of the multistage nature of cancer. In: *The toxicology of aflatoxins: Human health, veterinary, and agricultural significance*. Eaton DL, Groopman JD (eds). London: Academic Press. 1993, pp.179–206.
70. Gorelick NJ, Bruce RD, Hoseyni MS. Human risk assessment based on animal data inconsistencies and alternatives. In: *The toxicology of aflatoxins: Human health, veterinary, and agricultural significance*. Eaton DL, Groopman JD (eds). London: Academic Press. 1993, pp.508-11.
71. Henry SH, Bosch FX, Bowers JC. Aflatoxin, hepatitis and worldwide liver cancer risks. *Adv Exp Med Biol* 2002; 504: 229 –33.

72. Groopman JD. Molecular dosimetry methods for assessing human aflatoxin exposures. The toxicology of aflatoxins: Human health, veterinary and agricultural significance. Eaton D, Groopman JD (eds). London: Academic Press. 1993, pp.259-79.
73. Wang JS, Huang T, Su JJ. Hepatocellular carcinoma and aflatoxin exposure in Zhuqing Village, Fusui County, People's Republic of China. *Cancer Epidemiol Biomarkers Prev* 2001; 10: 143–6.
74. Sylla A, Diallo MS, Castegnaro JJ, Wild CP. Interactions between hepatitis B virus infection and exposure to aflatoxins in the development of hepatocellular carcinoma: A molecular epidemiological approach. *Mutat Res* 1999; 428: 187–96.
75. Hatch MC, Chen CJ, Levin B. Urinary aflatoxin levels, hepatitis-B virus infection and hepatocellular carcinoma in Taiwan. *Int J Cancer* 1993; 54: 931–4.
76. Groisman IJ, Koshy R, Henkler F, Groopman JD, Alaoui-Jamali MA. Down regulation of DNA excision repair by the hepatitis B virus-x protein occurs in p53-proficient and p53-deficient cells. *Carcinogenesis* 1999; 20: 479 – 83.
77. El-Zayadi AR, Badran HM, Barakat EMF, Attia MED, Shawky S, Mohamed MK, et al. Hepatocellular carcinoma in Egypt: A single center study over a decade. *World J. Gastroenterol* 2005; 11(33): 5193–8.
78. El-Kafrawy SA, Abdel-Hamid M, El-Daly M, Nada O, Ismail A, Ezzat S, et al. P53 mutations in hepatocellular carcinoma patients in Egypt. *Int J Hyg. Environ. Health* 2005; 208(4): 263–70.
79. Hifnawy MS, Mangoud AM, Eissa MH, Nor Edin E, MostafaY, Abou el Magd Y. The role of aflatoxin-contaminated food materials and HCV in developing hepatocellular carcinoma in Al-Sharkia Governorate, Egypt. *J. Egypt Soc. Parasitol* 2004; 34(1): 479–88.
80. Gong YY, Cardwell K, Hounsa A, Egal S, Turner PC, Hall AJ, et al. Dietary aflatoxin exposure and impaired growth in young children from Benin and Togo: Cross sectional study. *BMJ* 2002; 325: 20-1.
81. Maxwell SM, Apeagyei F, Vries HR, Mwanmut DD, Hendrickse RG. Aflatoxins in breast milk, neonatal cord blood and sera of pregnant women. *J Toxicol Toxin Rev* 1989; 8(1-2): 19-29.
82. Polychronaki N, Turner CP, Mykkanen H, Gong Y, Amra H, Abdel Wahhab M, et al. Determinants of aflatoxin M₁ in breast milk in a selected group of Egyptian mothers, *Food Addit Contam* 2006; 23(7):700-8.
83. Hassan AM, Sheashaa HA, Abdel Fatah MF, Ibrahim AZ, Gaber OA. Does aflatoxin as an environmental mycotoxin adversely affect the renal and hepatic functions of Egyptian lactating mothers and their infants? A preliminary report. *Int Urol Nephrol* 2006; 38(2): 339–42.

84. Paulín EGL, Martínez EM, Castro SPM. Aflatoxins and their impact on human and animal health: An emerging problem. In: Aflatoxins biochemistry and molecular biology. Gonzalez RGG (ed). In Tech, DOI. 2011, pp.265-9.
85. Verma RJ. Aflatoxins cause DNA damage. *Int J Hum Genet* 2004; 4(4): 231- 6.
86. Patterson DSP. Metabolism as a Factor in determining the toxic action of the aflatoxins in different animal species. *Food Cosmet Toxicol* 1973; 11: 287-94.
87. Food and Agriculture Organization of the United Nations (FAO). Worldwide regulations for mycotoxins in food and feeds in 2003. Food and Nutrition Paper No 81. Rome, Italy: FAO; 2003.
88. Wild CP, Gong YY. Mycotoxins and human disease: A largely ignored global health issue. *Carcinogenesis* 2010; 31(1): 71-82.
89. Copetti MV, Iamanaka BT, Pereira JL, Lemes DP, Nakano F, Taniwaki MH. Co-occurrence of ochratoxin A and aflatoxins in chocolate marketed in Brazil. *Food Control* 2012; 26(1): 36-41.
90. Dowd PF. Insect management to facilitate preharvest mycotoxin management. *Toxins* 2003; 22(2-3): 327-50.
91. Bruns HA. Controlling aflatoxin and fumonisin in maize by crop management. *Toxin Rev* 2003; 22(2-3): 153-73.
92. Hell K, Mutegi C. Aflatoxin control and prevention strategies in key crops of Sub-Saharan Africa. *Afr J Microbiol Res* 2011; 5(5): 459-66.
93. Diener UL, Cole RJ, Sanders TH, Payne GA, Lee LS, Klich MA. Epidemiology of aflatoxin formation by *Aspergillus flavus*. *Annu Rev Phytopathol* 1987; 25: 240.
94. Bolet AMB, Socarrás SMM. Micotoxinas y cáncer. *Revista Cubana de Investigaciones Biomédicas* 2005; 24(1): 54-9.
95. Food and Agriculture Organization of the United Nations (FAO). Control of aflatoxin in maize. In: Agriculture and Consumer Protection (ed.) Mycotoxin prevention and control in foodgrains. Rome, Italy: FAO; 1989.
96. Borrell J, Gimeno G. Micotoxinas en los alimentos: Medidas de prevención y detoxificación. *Selecciones Avícolas* 2002; 1: 567-71.
97. Varma J, Dubey N. Efficacy of essential oils of *Caesulia axillaris* and *Mentha arensis* against some storage pests causing bio-deterioration of food commodities. *Int J Food Microbiol* 2001; 68(3): 207-10.
98. Gowda NKS, Malathi V, Suganthi RU. Effect of some chemical and herbal compounds on growth of *Aspergillus parasiticus* and aflatoxin production. *Anim Feed Sci Tech* 2004; 116(3-4): 281-91.

99. Fandohan P, Zoumenou D, Hounhouigan DJ, Marasas WF, Wingfield MJ, Hell K. Fate of aflatoxins and fumonisins during the processing of maize into food products in Benin. *Int J Food Microbiol* 2005; 98(3): 249-59.
100. Miller RB. Electronic radiation of foods. Springer Science and Business Media, Inc. 2005, pp.1-42.
101. Siwela AH, Siwela M, Matindi G, Dube S, Nziramasanga N. Decontamination of aflatoxin-contaminated maize by dehulling. *J Sci of Food Agr* 2005; 85(15): 2535-8.
102. Jay JM. Radiation protection of foods and nature of microbial radiation resistance. In: *Modern food microbiology*. 7th Ed. Springer Science and Business Media, Inc. 2005, pp.371- 94.
103. Loaharanu P, Ahmed M. Advantage and disadvantage of the use of irradiation for food preservation. *J Agr Environ Ethics* 1991; 4(1):14-30.
104. Rosentha I. Ionizing radiation. In: *Electronic radiation in food science*. Springer-Verlage Berlin Heidelberg. 1992, pp.9-64.
105. Aziz NH, Moussa LAA, El-Far FM. Reduction of fungi and mycotoxins formation in seeds by gamma-radiation. *Food Safety* 2004; 24(2): 109-27.
106. Aziz NH, EL-Far FM, Shahin AAM, Roushdy SM. Control of *Fusarium* moulds and fumonisin B₁ in seeds by gamma-irradiation. *Food Control* 2007; 18: 1337- 42.
107. Cișmileanu A, Voicu G, Ciucă V, Ionescu M. Determination of aflatoxin B₁ in cereal-based feed by a high-performance chromatographic method. *Lucrări Științifice Medicină Veterinară* 2008; XLI: 565-9.
108. Nielsen S. *Food analysis laboratory manual*. 2th ed. Springer New York Dordrecht Heidelberg London. 2010, p. 20(moisture), 31-2(fat) , 41-3 (protein) , 161-2(fatty acid).
109. Marshall MR. Ash content. In: *Food analysis laboratory manual*. Nielsen S (ed). 4th Ed. Springer New York Dordrecht Heidelberg London. 2010, pp.109-10.
110. Egan H, Kirk RS, Sawyer R. *Pearson chemical analysis of food*. 8th Ed. Churchill Livingstone New York. 1981, pp.4-5.
111. Amagloh FK, Hardacre A, Mutukumira AN, Weber JL, Brough L, Coad J. Sweet potato-based complementary food for infants in low-income countries. *Food Nutr Bull* 2012; 33: 3–10.
112. Anjum FM, Ahmad I, Butt MS, Sheikh MA, Pasha I. Amino acid composition of spring wheat and losses of lysine during chapatti baking. *J Food Comp Anal* 2005; 18: 523-32.
113. Felicia Wu, Hasan G. Aflatoxin regulations in a network of global maize trade. *PLoS One*.2012; 7(9): 4515.

114. Aziz NH, Youssef BM. Inactivation of naturally occurring mycotoxins in some Egyptian foods and agricultural commodities by gamma-irradiation. *Egypt J Food Sci* 2002; 30: 167–77.
115. Ghanem I, Orfi M, Shamma M. Effect of gamma radiation on the inactivation of aflatoxin B₁ in food and feed crops. *Braz Journal Microbiol* 2008; 39: 787-91.
116. Stefanova R, Vasilev NV, Spassov SL. Irradiation of food, current legislation framework, and detection of irradiated foods. *Food Anal. Methods* 2010; 3: 225–52.
117. Stewart EM. Detection methods for irradiated foods. In: *Food irradiation principles and applications*. Molins R. (ed). Wiley Interscience. 2001, pp.347–86.
118. Gomes HA, Silva EN, Cardello HM, Cipolli KM. Effect of gamma irradiation on refrigerated mechanically deboned chicken meat quality. *Meat Sci* 2003; 65: 919–26.
119. Stefanova R. Effect of gamma-ray irradiation on the fatty acid profile of irradiated beef meat. *Food Chem* 2011; 127(2): 461-6.
120. Ahsan S, Hussain Z, Naqvi SA, ASI MR. Effect of gamma radiation on aflatoxin load, amino acid and fatty acid composition of *Oryza sativa l*. *Pak. J. Bot* 2013; 45(5): 1577-80.
121. Siddhuraju P, Osoniyi O, Makkar HPS, Becker K, Effect of soaking and ionizing radiation on various antinutritional factors of seeds from different species of an unconventional legume, *Sesbania* and a common legume, green gram (*Vigna radiata*). *Food Chem* 2002; 79: 273-81.
122. World Health Organization (WHO). Wholesomeness of irradiated food. Technical Report Series, No. 604. Geneva: WHO; 1977.
123. Satter A, Neelofar MA. Irradiation and germination effects on phytate, protein and amino acids of soybean. *Plant Food Hum Nutr* 1990; 40: 185-95.
124. Joseph OA, Klasus M, Kwaku D, Amanda M. Functional properties of cowpea (*Vigna unguiculata* L.Walp) flours and pastes as affected by gamma irradiation. *Food Chem* 2005; 93: 103-11.
125. World Health Organization (WHO). High dose irradiation: Wholesomeness of food irradiated with dose above 10 kGy, technical report series 890. Geneva: WHO; 1994.

الملخص العربي

يعتبر حدوث السموم الفطرية في المواد الغذائية (الحبوب) أمر لا مفر منه، و من التحديات المصاحبة لإنتاج المحاصيل فقدان حوالي ٢٥% من المحاصيل سنويا بسبب السموم الفطرية على حسب ما اعلنته منظمة الاغذية والزراعة.

الافلاتوكسن ب١ من اخطر السموم الفطرية المعروفة في الحيوانات ومصنف على حسب الوكالة الدولية لأبحاث السرطان كمجموعة أولى مسرطنة وهذا يعني أنه من العوامل التي تحفز السرطان التي أثبتت جدواها، و الأفلاتوكسين ب١ هي مجموعة من السموم الفطرية المنتجة في المناطق الاستوائية وشبه الاستوائية، و الأفلاتوكسين ب١ ناتج من نواتج الايض الثانوية لفطر الاسبرجلس فلافس و الاسبرجلس براسيكس ويلوث عدد كبير من المحاصيل في البلدان ذات المناخ الحار والرطب.

إن الهدف من هذه الدراسة هو تقييم التأثير المحتمل لاشعة جاما على الحد من الافلاتوكسن ب١ وتأثير ذلك على القيم الغذائية بما في ذلك الرماد، والرطوبة، والدهون، والبروتين، والكربوهيدرات، والالياف الخام، والسعرات الحرارية، والأحماض الدهنية، والأمنية وقد اجريت الدراسة على عدد ستين عينة موزعة بالتساوي على ثلاث حبوب وهى الذرة والقمح والأرز ولتحقيق هذا الهدف تم تنفيذ ما يلي :-

لقد تم اختيار عينات عشوائية من الأسواق المحلية وأسواق الجملة في مدينة الإسكندرية في شهر يونيه ويوليو واغسطس ٢٠١٣، و تم تقسيم كل عينة (واحد كيلوجرام) إلى اقسام متساوية لأربع مجموعات فرعية، تم تخزين العينات في أكياس بلاستيكية في درجة تبريد اربع درجات حتى يتم التحليل.

اولا- تم أخذ الربع الأول من كل عينات الحبوب الفرعية (العينة الضابطة) إلى المختبر المركزي للمعهد العالي للصحة العامة لاجراء التحاليل التالية:

- ١- تحديد نسبة الأفلاتوكسين ب١ بجهاز الكرماتوجرافيا السائل عالي الكفاءة.
 - ٢- تحليل محتوى الحبوب من المكونات الغذائية ويشمل:
 - تحديد محتوى الرطوبة
 - تحديد محتوى الدهن.
 - تقدير محتوى البروتين.
 - تحديد محتوى الرماد.
 - تحديد نسبة الالياف الخام
 - حساب محتوى الكربوهيدرات.
 - حساب السعرات الحرارية.
 - ٣- تحضير استرات الميثيل للأحماض الدهنية بطريقة الصوديوم ميثوكسيد ثم تحليل الأحماض الدهنية بواسطة جهاز كروماتوجرافيا الغاز.
 - ٤- تحليل الأحماض الأمينية بواسطة جهاز محلل الأحماض الأمينية.
- ثانيا- تم تعريض الربع الثاني من العينة من كل نوع من الحبوب للاشعاع باستخدام مصدر أشعة جاما المنبعثة من الكوبلت-٦٠ عند جرعة ٤ كيلو جرای ثم اجريت لها نفس التحاليل السابقة.
- ثالثا- تم تعريض الربع الثالث من العينة من كل نوع من الحبوب للاشعاع باستخدام مصدر أشعة جاما المنبعثة من الكوبلت-٦٠ عند جرعة ٦ كيلو جرای ثم اجريت لها نفس التحاليل السابقة.

رابعاً- تم تعريض الربع الرابع من العينة من كل نوع من الحبوب للإشعاع باستخدام مصدر أشعة جاما المنبعثة من الكوبلت-٦٠ عند جرعة ٨ كيلو جراى ثم اجريت لها نفس التحاليل السابقة الذكر.

وقد تم تحليل نتائج هذه الدراسة إحصائياً و تبين ما يلي:

- ١- عينات الذرة تحتوي على أعلى مستوى من الأفلاتوكسين أكثر من القمح والأرز.
- ٢- أشعة جاما تقلل من مستويات الأفلاتوكسين ب١ في عينات الحبوب دون التأثير على القيم الغذائية.
- ٣- أشعة جاما عند ٤ كيلو جراى، قللت من نسبة افلاتوكسين ب١ بمعدل ١٥.٥٤%، و % ٢٢.٢٥ و ٢٧.٤٦، فى الذرة والقمح والأرز على التوالي، وعند ٦ كيلو جراى قللت من افلاتوكسين ب١ بمعدل ٣٢.٣٩%، و ٤٣.٨٤، و ٥٦.٣٨% فى الذرة والقمح والأرز على التوالي. وجرعة الاشعاع ٨ كيلو جراى إزالت حوالي ٦٠.٢٦% من السم فى الذرة، ٦٤.٦٨% فى الأرز، ٦٩.٢٩% فى القمح.
- ٤- وبناء على هذه النتائج يفضل استخدام الجرعات الإشعاعية اعلى من ٨ كيلو جراى حتى يصل السم للمعدل المسموح به قانوناً على حسب ما اعلنته منظمة الاغذية والزراعة (فاو).
- ٥- لقد بينت الدراسة ان زيادة نسبة الزيوت فى الحبوب يكون عاملاً من عوامل انخفاض نسبة الاقلال من افلاتكسين ب١، حيث ان اقل انخفاض بواسطة الاشعاع كان فى حبوب الذرة نظراً لاحتوائه على محتوى اعلى من الزيوت وهذا ما تم تاييده فى دراسات أخرى.
- ٦- تؤثر أشعة جاما على بعض الأحماض الدهنية فى القمح والأرز، حيث تسبب انخفاضاً فى الحمض الدهني الاحادي غير المشبع (حامض الأوليك) و الحمض الدهني العبيد غير المشبع (حمض اللينولينيك) الذى أختفى نهائياً فى حبوب الارز عند جرعة الاشعاع ٨ كيلو جراى ، اما فى الذرة فحدثت زيادة فى الحمض الدهنى البلمتك ونقص فى الحمض الدهني الاحادي غير المشبع (حامض الأوليك)، وهذا ينطبق مع العديد من الدراسات.
- ٧- تؤثر أشعة جاما قليلاً على الأحماض الأمينية فى الحبوب، فى حبوب الذرة حدثت زيادة فى الليوسين اما الفينيل ألانين والثريومين فيقل كل منهما، اما حبوب القمح فحدثت انخفاض فى كل من التربتوفان، والايزولويسين، والفنيل ألانين، والارجنين، والليسين، وزيادة فى الليوسين اما حبوب الارز فحدثت زيادة فى الليوسين وانخفاض فى الفينيل ألانين ويختلف مقدار الزيادة والنقصان باختلاف نوع الحبوب وجرعة الاشعاع.



جامعة الإسكندرية
معهد البحوث الطبية
قسم الفيزياء الحيوية الطبية

تقييم التأثير المحتمل لاشعة جاما على الحد من افلاتكسن ب₁ فى بعض الحبوب وأثر ذلك على القيم الغذائية

رسالة مقدمة

لقسم الفيزياء الحيوية الطبية - معهد البحوث الطبية - جامعة الإسكندرية
ضمن متطلبات درجة

الماجستير

فى

الفيزياء الحيوية الطبية

من

عايدة صابر حمودة احمد

بكالوريوس العلوم (كيمياء حيوية خاصة) ، ١٩٩٦ ،
كلية العلوم ، جامعة الإسكندرية

[أكتوبر/ ٢٠١٤]



جامعة الإسكندرية
معهد البحوث الطبية
قسم الفيزياء الحيوية الطبية

تقييم التأثير المحتمل لاشعة جاما على الحد من افلاتكسن ب₁ فى بعض الحبوب وأثر ذلك على القيم الغذائية

رسالة مقدمة من

عايدة صابر حمودة احمد

للحصول على درجة

الماجستير

فى

الفيزياء الحيوية الطبية

التوقيع

.....

.....

.....

.....

لجنة المناقشة والحكم على الرسالة

أ.د.حسين على محمد مطاوع

أستاذ ورئيس قسم الفيزياء
كلية العلوم - جامعة دمنهور

أ.د.متولى على متولى قطب

أستاذ الفيزياء الحيوية الطبية
معهد البحوث الطبية - جامعة الإسكندرية

أ.د.سهير محمود الخولى

أستاذ ورئيس قسم الفيزياء الحيوية الطبية
معهد البحوث الطبية - جامعة الإسكندرية

د. نيفين فهمى محمد عجمى

أستاذ مساعد بقسم التغذية
المعهد العالى للصحة العامة- جامعة الإسكندرية

لجنة الإشراف

موافقون

الدكتور/ متولى على متولى قطب

أستاذ متفرغ الفيزياء الحيوية الطبية
قسم الفيزياء الحيوية الطبية
معهد البحوث الطبية
جامعة الإسكندرية

.....

الدكتورة/ نيفين فهمى محمد عجمى

أستاذ مساعد بقسم التغذية
قسم التغذية
المعهد العالى للصحة العامة
جامعة الإسكندرية

.....

الدكتورة/ رشا سعيد شمس الدين

زميل بقسم الفيزياء الحيوية الطبية
قسم الفيزياء الحيوية الطبية
معهد البحوث الطبية
جامعة الإسكندرية

.....