

# **RECOMMENDATIONS**

- 1- Human leukocyte antigen- G should be done on a wider scale of AML patients to confirm its relation to unfavourable risk patients.
- 2- Further studies should be performed to assess HLA-G role in context with other genetic and epigenetic factors that affect the prognosis of AML patients.

## **BIBLIOGRAPHY**

1. Smith BD, Sung L. Acute myeloid leukemia. *blood*. 2013;5(18):481-9.
2. Vardiman JW, Harris NL, Brunning RD. The World Health Organization (WHO) classification of the myeloid neoplasms. *Blood*. 2002;100(7):2292-302.
3. Baer MR, Greer JP. Acute Myeloid Leukemia in adults. In: Greer J, Foerster J, Lukens JN (eds). *Wintrobe's Clinical Hematology*. Lippincott: Williams & Wilkins Publisher. 12<sup>th</sup> edition. 2009;1920-32.
4. Suhag VS, Malkovska V, Solomon SR. Acute Myeloid leukemia. In: Rodgers GP, Young NS, eds. *Bethesda Handbook of Clinical Hematology*. Lippincott Williams & Wilkins. 3<sup>rd</sup> edition. 2013;131-47.
5. Welch John S, Ley Timothy J, Link Daniel C, Miller Christopher A, Larson David E, Koboldt Daniel C, et al. The Origin and Evolution of Mutations in Acute Myeloid Leukemia. *Cell* 2012;150(2):264-78.
6. Marcucci G, Haferlach T, Dohner H. Molecular genetics of adult acute myeloid leukemia: prognostic and therapeutic implications. *J Clin Oncol* 2011;29(5):475-86.
7. Frohling S, Scholl C, Gilliland DG, Levine RL. Genetics of myeloid malignancies: pathogenetic and clinical implications. *J Clin Oncol* 2005;23(26):6285-95.
8. Welch JS. Subclonal architecture in acute myeloid leukemia. *Hematology: the education program for the annual congress of the European Hematology Association*. 2013;7:23-9.
9. Cagnetta A, Adamia S, Acharya C, Patrone F, Miglino M, Nencioni A, et al. Role of genotype-based approach in the clinical management of adult acute myeloid leukemia with normal cytogenetics. *Leukemia Research* 2014;38(6):649-59.
10. Sangle NA, Perkins SL. Core-binding factor acute myeloid leukemia. *Archives of pathology & laboratory medicine*. 2011;135(11):1504-9.
11. Chen T, Jiang G, Fu X, Wang L, Qian H, Wu K, et al. CK19 mRNA expression measured by reverse-transcription polymerase chain reaction (RT-PCR) in

the peripheral blood of patients with non-small cell lung cancer treated by chemo-radiation: an independent prognostic factor. *Lung Cancer* 2007;56:105 - 14.

12. Moldenhauer A, Frank RC, Pinilla-Ibarz J, Holland G, Boccuni P, Scheinberg DA, et al. Histone deacetylase inhibition improves dendritic cell differentiation of leukemic blasts with AML1-containing fusion proteins. *Journal of Leukocyte Biology* 2004;76(3):623-33.

13. Faridi F, Ponnusamy K, Quagliano-Lo Coco I, Chen-Wichmann L, Grez M, Henschler R, et al. Aberrant epigenetic regulators control expansion of human CD34+ hematopoietic stem/progenitor cells. *Frontiers in genetics* 2013;4:254.

14. Paschka P, Dohner K. Core-binding factor acute myeloid leukemia: can we improve on HiDAC consolidation? *Hematology*. 2013:209-19.

15. Fröhling S, Schlenk RF, Kayser S, Morhardt M, Benner A, Döhner K, et al. Cytogenetics and age are major determinants of outcome in intensively treated acute myeloid leukemia patients older than 60 years: results from AMLSG trial AML HD98-B. *Blood* 2006;108(10):3280-8.

16. Chen J, Odenike O, Rowley JD. Leukaemogenesis: more than mutant genes. *Nat Rev Cancer* 2010;10(1):23-36.

17. Ismael O, Shimada A, Hama A, Elshazley M, Muramatsu H, Goto A, et al. De novo childhood myelodysplastic/myeloproliferative disease with unique molecular characteristics. *Br J Haematol* 2012;158(1):129-37.

18. Bullinger L, Döhner H. Genetics guided therapeutic approaches in acute myeloid leukemia. *European Hematology Association education book*. 2013;7:30-40.

19. Appelbaum FR, Kopecky KJ, Tallman MS, Slovak ML, Gundacker HM, Kim HT, et al. The clinical spectrum of adult acute myeloid leukaemia associated with core binding factor translocations. *Br J Haematol*. 2006;135(2):165-73.

20. Park JH, Qiao B, Panageas KS, Schymura MJ, Jurcic JG, Rosenblat TL, et al. Early death rate in acute promyelocytic leukemia remains high despite all-trans retinoic acid. *Blood*. 2011;118(5):1248-54.

21. Bernasconi P, Dambruoso I, Boni M, Astori C, Cavigliano PM, Zappasodi P, et al. Chromosomal abnormalities and TET2 involvement in therapy-related myelodysplastic syndromes (t-MDS) and acute myeloid leukemias (t-AML). *ASH Annual Meeting Abstracts*. 2009.

22. Dohner H, Gaidzik VI. Impact of genetic features on treatment decisions in AML. *Hematology Am Soc Hematol Educ Program* 2011:36-42.
23. Deneberg S, Guardiola P, Lennartsson A, Qu Y, Gaidzik V, Blanchet O, et al. Prognostic DNA methylation patterns in cytogenetically normal acute myeloid leukemia are predefined by stem cell chromatin marks. *Blood* 2011;118(20):5573-82.
24. Vassiliou GS, Cooper JL, Rad R, et al. Mutant nucleophosmin and cooperating pathways drive leukemia initiation and progression in mice. *Nat Genet.* 2011;43(5):470-5.
25. Lindström MS. NPM1/B23: A multifunctional chaperone in ribosome biogenesis and chromatin remodeling. *Biochem Res Int* 2011;2011: 195209.
26. Cornelissen JJ, van Putten WL, Verdonck LF, Theobald M, Jacky E, Daenen SM, et al. Results of a HOVON/SAKK donor versus no-donor analysis of myeloablative HLA-identical sibling stem cell transplantation in first remission acute myeloid leukemia in young and middle-aged adults: benefits for whom? *Blood.* 2007;109(9):3658-66.
27. Metzeler KH, Dufour A, Benthaus T, Hummel M, Sauerland M-C, Heinecke A, et al. ERG expression is an independent prognostic factor and allows refined risk stratification in cytogenetically normal acute myeloid leukemia: a comprehensive analysis of ERG, MN1, and BAALC transcript levels using oligonucleotide microarrays. *J Clin Oncol* 2009;27(30):5031-8.
28. Schlenk RF, Dohner K, Krauter J, et al. Mutations and treatment outcome in cytogenetically normal acute myeloid leukemia. *N Engl J Med* 2008;358(18):1909-18.
29. Kronke J, Schlenk RF, Jensen KO, et al. Monitoring of minimal residual disease in NPM1 mutated acute myeloid leukemia: A study of the German-Austrian AML Study Group (AMLSSG). *J Clin Oncol* 2011;29(19):2709-16.
30. Schlenk RF, Dohner K, Kneba M, et al. Gene mutations and response to treatment with all-trans retinoic acid in elderly patients with acute myeloid leukemia. Results from AMLSSG trial AML HD98B. *Haematologica* 2009;94(1):54-60.
31. Becker H, Marcucci G, Maharry K, et al. Favorable prognostic impact of NPM1 mutations in older patients with cytogenetically normal de novo acute myeloid

leukemia and associated gene- and microRNA-expression signatures: a Cancer and Leukemia Group B study. *J Clin Oncol* 2010;28(4):596-604.

32. Abdel-Wahab O, Mullally A, Hedvat C, Garcia-Manero G, Patel J, Wadleigh M, et al. Genetic characterization of TET1, TET2, and TET3 alterations in myeloid malignancies. *Blood* 2009;114(1):144-7.

33. Damm F, Kosmider O, Gelsi-Boyer V, Renneville A, Carbuccia N, Hidalgo-Curtis C, et al. Mutations affecting mRNA splicing define distinct clinical phenotypes and correlate with patient outcome in myelodysplastic syndromes. *Blood* 2012; 119(14): 3211-8.

34. Becker H, Maharry K, Radmacher MD, Mrózek K, Metzeler KH, Whitman SP, et al. Clinical outcome and gene- and microRNA-expression profiling according to the Wilms tumor 1 (WT1) single nucleotide polymorphism rs16754 in adult de novo cytogenetically normal acute myeloid leukemia: a Cancer and Leukemia Group B study. *Haematologica*. 2011;96(10):1488-95.

35. Arneson W, Brickell J. *Clinical Chemistry. A Laboratory Perspective*. Company. F. A. Davis Company; 2007.

36. Abdel-Wahab O, Manshouri T, Patel J, Harris K, Yao J, Hedvat C, et al. Genetic analysis of transforming events that convert chronic myeloproliferative neoplasms to leukemias. *Cancer Res* 2010;70(2):447-52.

37. Grimwade D, Hills RK. Independent prognostic factors for AML outcome. *ASH Education Program Book* 2009;2009(1):385-95.

38. Rucker FG, Schlenk RF, Bullinger L, Kayser S, Teleanu V, Kett H, et al. TP53 alterations in acute myeloid leukemia with complex karyotype correlate with specific copy number alterations, monosomal karyotype, and dismal outcome. *Blood* 2012;119(9):2114-21.

39. Testa JR, Mintz U, Rowley JD, Vardiman JW, Golomb HM. Evolution of karyotypes in acute nonlymphocytic leukemia. *Cancer Res* 1997 ;39(9):3619-27.

40. Anderson K, Lutz C, van Delft FW, Bateman CM, Guo Y, Colman SM, et al. Genetic variegation of clonal architecture and propagating cells in leukaemia. *Nature* 2011;469(7330):356-61.

41. Schmetzger HM, Gerhartz HH. Acute myeloid leukemia (AML) can be oligoclonal. *Leukemia* 1993;7(12):1965-70.

42. Lo Coco F, Saglio G. Single or multistep origin of hemopoietic tumors: the contribution of clonality studies. *Leukemia* 1995;9(9):1586-9.
43. Estey EH. Acute myeloid leukemia: 2012 update on diagnosis, risk stratification, and management. *American journal of hematology* 2012;87(1):89-99.
44. Kohlmann A, Grossmann V, Nadarajah N, Haferlach T. Next generation sequencing-feasibility and practicality in haematology. *Br J Haematol* 2013;160:736-53.
45. Delhommeau F, Dupont S, Valle VD, James C, Trannoy S, Massé A, et al. Mutation in TET2 in Myeloid Cancers. *New Engl J Med* 2009;360(22):2289-301.
46. Kovats S, Main EK, Librach C, Stubblebine M, Fisher SJ, DeMars R. A class I antigen, HLA-G, expressed in human trophoblasts. *Science* 1990;248(4952):220-3.
47. Jurisicova A, Casper RF, MacLusky NJ, Librach CL. Embryonic human leukocyte antigen-G expression: possible implications for human preimplantation development. *Fertility and sterility* 1996;65(5):997-1002.
48. Fuzzi B, Rizzo R, Criscuoli L, Noci I, Melchiorri L, Scarselli B, et al. HLA-G expression in early embryos is a fundamental prerequisite for the obtainment of pregnancy. *European journal of immunology* 2002;32(2):311-5.
49. Hviid TV. HLA-G in human reproduction: aspects of genetics, function and pregnancy complications. *Human reproduction update*. 2006;12(3):209-32.
50. Rouas-Freiss N, Goncalves RM, Menier C, Dausset J, Carosella ED. Direct evidence to support the role of HLA-G in protecting the fetus from maternal uterine natural killer cytotoxicity. *Proceedings of the National Academy of Sciences of the United States of America*. 1997;94(21):11520-5.
51. Crisa L, McMaster MT, Ishii JK, Fisher SJ, Salomon DR. Identification of a thymic epithelial cell subset sharing expression of the class Ib HLA-G molecule with fetal trophoblasts. *The Journal of experimental medicine*. 1997;186(2):289-98.
52. Le Discorde M, Moreau P, Sabatier P, Legeais JM, Carosella ED. Expression of HLA-G in human cornea, an immune-privileged tissue. *Human immunology*. 2003;64(11):1039-44.
53. Cirulli V, Zalatan J, McMaster M, Prinsen R, Salomon DR, Ricordi C, et al. The class I HLA repertoire of pancreatic islets comprises the nonclassical class Ib antigen HLA-G. *Diabetes* 2006;55(5):1214-22.

54. Menier C, Rabreau M, Challier JC, Le Discorde M, Carosella ED, Rouas-Freiss N. Erythroblasts secrete the nonclassical HLA-G molecule from primitive to definitive hematopoiesis. *Blood* 2004;104(10):3153-60.
55. Menier C, Guillard C, Cassinat B, Carosella ED, Rouas-Freiss N. HLA-G turns off erythropoietin receptor signaling through JAK2 and JAK2 V617F dephosphorylation: clinical relevance in polycythemia vera. *Leukemia*. 2008;22(3):578-84.
56. Carosella ED, Moreau P, Le Maoult J, Le Discorde M, Dausset J, Rouas-Freiss N. HLA-G molecules: from maternal-fetal tolerance to tissue acceptance. *Advances in immunology* 2003;81:199-252.
57. Favier B, LeMaoult J, Rouas-Freiss N, Moreau P, Menier C, Carosella ED. Research on HLA-G: an update. *Tissue Antigens* 2007;69(3):207-11.
58. Rouas-Freiss N, Naji A, Durrbach A, Carosella ED. Tolerogenic functions of human leukocyte antigen G: from pregnancy to organ and cell transplantation. *Transplantation* 2007;84(Suppl 1):S21-5.
59. Rouas-Freiss N, Moreau P, Menier C, LeMaoult J, Carosella ED. Expression of tolerogenic HLA-G molecules in cancer prevents antitumor responses. *Semin Cancer Biol* 2007;17(6):413-21.
60. Dunn GP, Old LJ, Rd S. The three Es of cancer immunoediting. *Annu Rev Immunol* 2004;22:329-60.
61. Burnet M. Cancer; a biological approach. I. The processes of control. *Br Med J* 1957;6(5022):779-86.
62. Menier C, Rouas-Freiss N, Carosella ED. The HLA-G non classical MHC class I molecule is expressed in cancer with poor prognosis. Implications in tumour escape from immune system and clinical applications. *Atlas of Genetics and Cytogenetics in Oncology and Haematology*. 2008
63. Ishitani A, De G. Alternative splicing of HLA-G transcripts yields proteins with primary structures resembling both class I and class II antigens. *Proceedings of the National Academy of Sciences of the United States of America*. 1992;89(9):3947-51.
64. Kirszenbaum M, Moreau P, Gluckman E, Dausset JEC. An alternatively spliced form of HLA-G mRNA in human trophoblasts and evidence for the presence

of HLA-G transcript in adult lymphocytes. *Proceedings of the National Academy of Sciences of the United States of America* 1994;91(10):4209-13.

65. Paul P, Rouas-Freiss N, Khalil-Daher I, Moreau P, Riteau B, Le Gal FA, et al. HLA-G expression in melanoma: a way for tumor cells to escape from immunosurveillance. *Proceedings of the National Academy of Sciences of the United States of America*. 1998;95(8):4510-5.

66. Clements CS, Kjer-Nielsen L, McCluskey J, Rossjohn J. Structural Studies on HLA-G: Implications for Ligand and Receptor Binding. *Human immunology*. 2007;68:220-6.

67. Menier C, Saez B, Horejsi V, Martinozzi S, Krawice-Radanne I, Bruel S, et al. Characterization of monoclonal antibodies recognizing HLA-G or HLA-E: new tools to analyze the expression of nonclassical HLA class I molecules. *Human immunology* 2003;64(3):315-26.

68. Khalil-Daher I, Riteau B, Menier C, Sedlik C, Paul P, Dausset J, et al. Role of HLA-G versus HLA-E on NK function: HLA-G is able to inhibit NK cytotoxicity by itself. *J Reprod Immunol* 1999;43(2):175-82.

69. Riteau B, Menier C, Khalil-Daher I, Martinozzi S, Pla M, Dausset J, et al. HLA-G1 co-expression boosts the HLA class I-mediated NK lysis inhibition. *Int Immunol* 2001;13(2):193-201.

70. Selmani Z, Naji A, Zidi I, Favier B, Gaiffe E, Obert L, et al. Human leukocyte antigen-G5 secretion by human mesenchymal stem cells is required to suppress T lymphocyte and natural killer function and to induce CD4<sup>+</sup>CD25<sup>high</sup>FOXP3<sup>+</sup> regulatory T cells. *Stem Cells* 2007;26(1):212-22.

71. Cantoni C, Verdiani S, Falco M, Pessino A, Cilli M, Conte R, et al. p49, a putative HLA class I-specific inhibitory NK receptor belonging to the immunoglobulin superfamily. *European journal of immunology* 1998 ;28(6):1980-90.

72. Colonna M, Samaridis J, Cella M, Angman L, Allen RL, O'Callaghan CA, et al. Human myelomonocytic cells express an inhibitory receptor for classical and nonclassical MHC class I molecules. *J Immunol* 1998;160(7):3096-100.

73. Colonna M, Navarro F, Bellon T, Llano M, Garcia P, Samaridis J, et al. A common inhibitory receptor for major histocompatibility complex class I molecules

on human lymphoid and myelomonocytic cells. *The Journal of experimental medicine* 1997;186(11):1809-18.

74. Rajagopalan S, Long EO. A human histocompatibility leukocyte antigen (HLA)-G-specific receptor expressed on all natural killer cells. *The Journal of experimental medicine*. 1999 ;189(7):1093-100.

75. Shiroishi M, Tsumoto K, Amano K, Shirakihara Y, Colonna M, Braud VM, et al. Human inhibitory receptors Ig-like transcript 2 (ILT2) and ILT4 compete with CD8 for MHC class I binding and bind preferentially to HLA-G. *Proceedings of the National Academy of Sciences of the United States of America*. 2003;100(15):8856-61.

76. Edgardo D. Carosella, Benoit Favier, Nathalie Rouas-Freiss, LeMaout J. Beyond the increasing complexity of the immunomodulatory HLA-G molecule. *blood* 2008;111:4862-70.

77. Boyson JE, Erskine R, Whitman MC, Chiu M, Lau JM, Koopman LA, et al. Disulfide bond-mediated dimerization of HLA-G on the cell surface. *Proceedings of the National Academy of Sciences of the United States of America* 2002;99(25):16180-5.

78. Apps R, Gardner L, Sharkey AM, Holmes NAM. A homodimeric complex of HLA-G on normal trophoblast cells modulates antigen-presenting cells via LILRB1. *European journal of immunology* 2007;37(7):1924-37.

79. Shiroishi M, Kuroki K, Rasubala L, Tsumoto K, Kumagai I, Kurimoto E, et al. Structural basis for recognition of the nonclassical MHC molecule HLA-G by the leukocyte Ig-like receptor B2 (LILRB2/LIR2/ILT4/CD85d). *Proceedings of the National Academy of Sciences of the United States of America*. 2006;103(44):16412-7.

80. Shiroishi M, Kuroki K, Ose T, Rasubala L, Shiratori I, Arase H, et al. Efficient leukocyte Ig-like receptor signaling and crystal structure of disulfide-linked HLA-G dimer. *The Journal of biological chemistry* 2006;281(15):10439-47.

81. Gonen-Gross T, Achdout H, Arnon TI, Gazit R, Stern N, Horejsi V, et al. The CD85J/leukocyte inhibitory receptor-1 distinguishes between conformed and beta 2-microglobulin-free HLA-G molecules. *J Immunol* 2005;175(8):4866-74.

82. Gonen-Gross T, Achdout H, Gazit R, Hanna J, Mizrahi S, Markel G, et al. Complexes of HLA-G protein on the cell surface are important for leukocyte Ig-like receptor-1 function. *J Immunol* 2003;171(3):1343-51.
83. Creput C, Le Friec G, Bahri R, Amiot L, Charpentier B, Carosella E, et al. Detection of HLA-G in serum and graft biopsy associated with fewer acute rejections following combined liver-kidney transplantation: possible implications for monitoring patients. *Human immunology*. 2003;64(11):1033-8.
84. Rebmann V, Lemaoult J, Rouas-Freiss N, Carosella ED, Hg W. Report of the Wet Workshop for quantification of Soluble HLA-G in Essen, 2004. *Hum Immunol* 2005;66(8):853-63.
85. Carosella ED, Moreau P, LeMaoult J, Rouas-Freiss N. HLA-G: from biology to clinical benefits. *Trends in Immunology* 29(3):125-32.
86. Maejima M, Fujii T, Kozuma S, Okai T, Shibata YT. Presence of HLA-G-expressing cells modulates the ability of peripheral blood mononuclear cells to release cytokines. *Am J Reprod Immunol* 1997;38(2):79-82.
87. Pistoia V, Morandi F, Wang X, Ferrone S. Soluble HLA-G: Are they clinically relevant? *Seminars in Cancer Biology* 2007;17(6):469-79.
88. Apps R, Gardner L, Moffett A. A critical look at HLA-G. *Trends in Immunology*.29(7):313-21.
89. Ferrone S, FM. M. Loss of HLA class I antigens by melanoma cells: molecular mechanisms, functional significance and clinical relevance. *Immunol Today* 1995;16(10):487-94.
90. Ibrahim EC, Aractingi S, Allory Y, Borrini F, Dupuy A, Duvillard P, et al. Analysis of HLA antigen expression in benign and malignant melanocytic lesions reveals that upregulation of HLA-G expression correlates with malignant transformation, high inflammatory infiltration and HLA-A1 genotype. *Int J Cancer* 2004;108(2):243-50.
91. Swansbury J. Cancer cytogenetics: methods and protocols; cytogenetic techniques for myeloid disorders. *Methods in Molecular Biology* 2003;220:43-57.
92. Locafaro G, Amodio G, Tomasoni D, Tresoldi C, Ciceri F, Gregori S. HLA-G expression on blasts and tolerogenic cells in patients affected by acute myeloid leukemia. *Journal of Immunology Research* 2014;2014:10.

93. Yan WH, Lin A, Chen BG, Luo WD, Dai MZ, Chen XJ, et al. Unfavourable clinical implications for HLA-G expression in acute myeloid leukaemia. *J Cell Mol Med* 2008;12(3):889-98.
94. Alkhouly N, Shehata I, Ahmed MB, Shehata H, Hassan S, Ibrahim T. HLA-G expression in acute lymphoblastic leukemia: a significant prognostic tumor biomarker. *Medical Oncology* 2013;30(1):1-9.
95. Mizuno S, Emi N, Kasai M, Ishitani A, Saito H. Aberrant expression of HLA-G antigen in interferon gamma-stimulated acute myelogenous leukaemia. *Br J Haematol* 2000 ;111(1):280-2.
96. Poláková KN, Křčová M, Kuba D, Russ G. Analysis of HLA-G expression in malignant hematopoietic cells from leukemia patients. *Leukemia research*. 2003;27(7):643-8.
97. Guo Q, Chen B, Ruan Y, Lin A, Yan W. HLA-G expression is irrelevant to prognosis in patients with acute myeloid leukemia. *Leukemia Research* 2011;35(10):1350-4.
98. Gros F, Sebti Y, de Guiber S, Branger B, Bernard M, Fauchet R, et al. Soluble HLA-G molecules are increased during acute leukemia, especially in subtypes affecting monocytic and lymphoid lineages'. *Neoplasia*. 2006;8(3):223-30.
99. Curigliano G, Criscitiello C, Gelao L, Goldhirsch A. Molecular pathways: human leukocyte antigen G (HLA-G). *Clinical Cancer Research* 2013;19(20):5564-71.

## SUMMARY

Acute myeloid leukemia (AML) is a clonal stem cell malignancy in which immature hematopoietic cells proliferate and accumulate in bone marrow, peripheral blood, and other tissues. This process results in inhibition of normal hematopoiesis, and the clinical features of bone marrow failure. AML accounts for 90% of all acute leukemias in adults.

Human leukocyte antigen G (HLA-G) is a non-classic major histocompatibility complex class I gene encoding for a protein showing restrictive distribution and lower polymorphism. Membrane-bound HLA-G isoforms are expressed by extravillous cytotrophoblast cells, fetal capillary endothelial cells, endovascular cells, and thymic epithelium cells. They can also be detected in tumoral pathologies such as melanomas, breast, renal, and lung carcinomas gliomas, and cutaneous lymphomas.

HLA-G has been suggested to provide tumor cells with an effective pathway to escape from anti-tumor immune responses and to be involved in promoting beneficial tolerance in several settings, such as autoimmunity and organ transplantation, and in contributing to detrimental tolerance in viral infections and cancer. Few studies had been performed on HLA-G expression in different types of leukemia patients; however, data are limited and conclusions remain controversial. In addition, this molecule might be a promising target for future immune therapeutic approaches based on its immune tolerant functions and its highly specific expression for malignant transformation.

The present study aimed at determining the role of soluble HLA-G in AML patients and to study its prognostic significance. The study was performed on 30 patients and 15 healthy controls of matched age and sex. The mean value of age in group A was  $48.5 \pm 16.98$  years, in group B was  $50.1 \pm 17.5$  years and in group C was  $49.8 \pm 19.5$  years. Males in group A were 7 (46.67%), in group B were 6 (40%) and in group C were 8(53.33%). Females in group A were 8(53.33%), in group B were 9(60%) and in group C were 7 (46.67%). M4 was the commonest subtype in both

groups (6 cases, 40%). The mean value of sHLA-G levels in group A was  $329.8 \pm 57.54$  ng/L, in group B was  $451.15 \pm 163.99$  ng/L and in group C was  $551.63 \pm 109.08$  ng/L. There was statistically significant difference in sHLA-G levels between the three studied groups ( $P < 0.05$ ), being significantly higher in relapsed than new AML cases. The cutoff value of sHLA-G was 368.84, this value showed sensitivity of 100.0% and specificity of 62.0%. There was no statistically significant difference between sHLA-G as regards age, gender and WBCs ( $P > 0.05$ ) while there was statistically significant difference between, sHLA-G and bone marrow blasts percentage ( $P < 0.05$ ). There was statistically insignificant difference between sHLA-G and different FAB subtypes with the highest mean values was observed in M6 ( $605.3 \pm 116.5$ ) while the lowest mean value was observed in M2 ( $365.2 \pm 98.9$ ). ( $P = 0.685$ ). There was statistically insignificant difference between sHLA-G and response to therapy ( $P = 0.158$ ).

## المخلص

تعتبر اللوكيميا الميلودية الحادة من السرطانات التي تنشأ من الخلايا الجذعية المسرطنة و تحتوي على خلايا بدائية لها القدرة على التكاثر و التراكم في نخاع العظم و في الدم و انسجه اخرى مما يؤدي الى نقص الخلايا الطبيعيه في نخاع العظم و ظهور اعراض فشل النخاع. و تمثل اللوكيميا الميلودية حوالى 90% من كل اللوكيميا الحادة في البالغين.

الانتيجين الملائم النسيجي-جي يعتبر من مجموعه المورثات 1 غير النمطية و ينتج بروتين ذو انتشار وتعدد شكلي محدود. وقد وجد الانتجين الملائم -جي على الزغب الخارجية لخلايا التروفوبلاست و الشعيرات الدموية الجينية و خلايا البطان الوريدي و خلايا الثابوسية الغشائية. و قد وجد ايضا في الخلايا السرطانية مثل خلايا الميلانوما و خلايا سرطان الثدي و الكلى و الرئة و اورام المخ و اورام الليمفاوية الجلدية.

و يشكل الجزئي للانتجين الملائم النسيجي وظائف مناعية مختلفة من خلال تثبيط عمل الخلايا المناعية و برغم من ان اهميته ظهرت من التقبل المناعي بين الام و الجنين الا انه ظهر كاحد العوامل التي تساعد خلايا السرطان من تجنب التفاعلات المناعية مما يؤدي الى تقبل خلايا السرطان او حتى الخلايا المصابة بالفروسات الى جانب ظهور اهميته مع امراض اخرى مثل الامراض المناعية و زرع الاعضاء. توجد ابحاث قليلة عن العلاقة بين الانتجين الملائم النسيجي-جي وانواع اللوكيميا المختلفة ولكن النتائج محدودة و متنوعة. ولكن قد يكون ذو فائدة مستقبلية فى استخدام العلاج المناعى فى علاج الاورام

وكان هذا البحث يهدف الى دراسة دور الانتجين الملائم النسيجي - جي في لوكيميا النقوى الحادة ودراسة قيمته التنبؤية وقد اشتملت الدراسة على 30 مريضا مصابين باللوكيميا النقوية الحادة و 15 عينة ضابطة متوافقه من حيث العمر و النوع. وكان متوسط الاعمار في المجموعه الاولى  $48.5 \pm 16.98$  سنة و المجموعه الثانية  $50.1 \pm 17.5$  بينما كان فى المجموعه الثالثة  $49.8 \pm 19.5$  سنة. و بلغت نسبة الاناث في المجموعه الاولى 53.3% و في الثانية 60% و الثالثة 46.67%. و لوحظ ان اكثر انواع اللوكيميا شيوعا كان M4. و لم يتواجد اختلاف بين المورثات بين مجموعه المرضى المنتكسة و حديثى التشخيص. وبلغ مستوى الانتجين الملائم النسيجي جي في المجموعه الاولى  $329.8 \pm 57.54$  ng/L و في المجموعه الثانية  $451.15 \pm 163.99$  ng/L و فى المجموعه الثالثة  $551.63 \pm 109.08$  ng/L. و قد وجد اختلاف ذو دلالة احصائية بين هذه القيم فى مجموعه المرضى المنتكسة و مجموعه حديثى التشخيص بالمقارنة بالمجموعه الضابطة ( $p < 0.05$ ).

واتضح من الدراسة ان الحد الفاصل لنسبة الانتجين كان عند القيمة 368.84 حيث بلغ مدى حساسية التشخيص نسبة 100% و الخصوصية 62%. و عند تحليل الانتجين و علاقته ببقية العوامل لوحظ الاتي: وجدت علاقة ذات دلالة احصائية بينه و بين نسبة الخلايا السرطانية في نخاع العظم ( $P < 0.05$ ). الا انه لم تتواجد اى علاقة بينه و بين سن المرضى أو جنس المريض أو عدد كرات الدم البيضاء في الدم ( $p > 0.05$ ). كما انه لم تتواجد اى علاقة بينه و نوع اللوكيميا النقوى الحادة وكان مستوى الانتجين أعلى فى النوع (605.3±116.5) M6 بينما

كانت اقل قيمة له فى النوع M2 (365.2±98.9) بالمقارنة بالانواع الاخرى (P =0.685) . كما انه لم تتواجد اى علاقه ذات دلالة احصائية بينه و بين مدى استجابته للعلاج (P=0.158).

دراسة مستوى الانتيجين الملائم النسيجي- جى المرسل عند مرضى

اللويميا الميلودية الحادة

رسالة مقدمة من

الطبيب/ أحمد عبدالحميد كسبر

إيفاء جزئيا للحصول على درجة

الماجستير في أمراض دم

كلية الطب

جامعة الإسكندرية

التوقيع

لجنة الحكم على الرسالة

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أ.د / نبيل أحمد الحلواني

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استشاري الباثولوجيا الاكلينيكية و الكيمائية

الاكاديمية الطبية العسكرية

دراسة مستوى الانتيجين الملائم النسيجي- جى المرسل عند مرضى

اللوكميا الميلودية الحادة

رسالة

مقدمة إلى كلية الطب- جامعة الإسكندرية

إيفاء جزئياً للحصول علي درجة

الماجستير في أمراض دم

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بكالوريوس الطب والجراحة

كلية الطب- جامعة الاسكندرية

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