

8. SUMMARY

Plant secondary metabolites provide a wealth of medicinally useful compounds and play a crucial role in modern and traditional medicine. In some cases the compound of interest is easily identifiable, and can often be synthetically produced. However, in many cases it is not easy to determine whether the medicinal benefits are produced by one or several compounds or if they have additive or synergistic effects. In these instances a synthetic product cannot be produced and medicinal preparations will rely on plant material collected from the wild, grown using conventional cultivation techniques, or produced using *in vitro* methods. *Echinacea spp.* (family Asteraceae) herbal medicines and dietary supplements are traditionally used as immunostimulants in the treatment of inflammatory and viral diseases. *Echinacea purpurea* (L.) is an important commercial species. A great deal of research has indicated the chemical composition of *Echinacea spp.*, including alkamides, caffeic acid derivatives (chicoric acid, caftaric acid and chlorogenic acid), and polysaccharides attracting claims of beneficial pharmacological activity. *Echinacea* is an example of plant species with medicinal activity that cannot be attributed to a single chemical, necessitating plant production to obtain the full spectrum of metabolite. Several compounds are thought to play significant roles in the efficacy of *Echinacea* products and are often used to ensure and test for quality. Thus developing artificial (controlled) culture systems with the aims of standardizing and improving production and marketing of medicinal species allows rapid propagation of plants selected for their active principles and improves the quality and quantity of natural pharmaceutical compounds. Accordingly, the current study aimed to establish plant cell lines capable of producing high yields of secondary compounds in cell suspension cultures as an alternative to conventional whole plant production to improve the productivity of locally cultivated *Echinacea purpurea* and to ensure that produced compounds are effective. The targeted group of compounds to be studied was the polyphenolics (caffeic acid derivatives). The long-term goal of our research is to aid in the development of effective therapeutic preparations of *Echinacea*, the goals of this particular project are

1. Establishment of callus cultures from explants isolated from plant material:

Callus induction is necessary, as the first step, in many tissue culture experiments. Callus is produced when the initial response of the tissues to a wound is followed by the external addition of growth regulators in an aseptic medium in order to maintain the rapid cell division response and sustain it indefinitely. Calli can be obtained from almost any part of the plant; here the root was chosen to be the explant. Phytohormones, such as cytokinin and auxin, are necessary ingredients in any plant cell culture environment. Generally speaking, an equal proportion of auxin to cytokinin hormones in solution will impact only cell proliferation and result in the formation of callus masses. In our study callus induction was achieved using MS media supplemented with 1.5mg/L BA and 0.5mg/L NAA.

2. Establishment of liquid cultures (suspension cultures) from static cultures:

To initiate mother cell cultures, pieces of established undifferentiated, friable calli were subdivided into small sections and transferred from solid media to liquid media supplemented with 1.5mg/L BA and 0.5mg/L NAA. Periodic addition of fresh media and draining out the exhausted media ensures maintenance of suspension cultures under steady state of growth for long periods. Cell suspension cultures were sub-cultured for several weeks to expand the total biomass to be collected.

3. Study of the influence of elicitors and precursor feeding on the accumulation of active principles in suspension cell cultures:

Production of secondary metabolites can be enhanced by the treatment of the undifferentiated cells with elicitors. An 'elicitor' may be defined as a substance which, when introduced in small concentrations to a Living cell system, initiates or improves the biosynthesis of specific compounds. Elicitation is the induced or enhanced biosynthesis of metabolites due to addition of trace amounts of elicitors. The elicitors used in this study were: MeJA, chitosan, SNP, CuSO₄ and. MeJA solution was used with concentrations 0, 5, 10 and 20 mg/L whereas chitosan used at concentrations 0, 5, 25 and 50 mg/L, SNP used concentrations were 0, 50, 100 and 250 μ M and CuSO₄ concentrations were 0, 2, 4 and 8 μ M, Also, the impact of 1, 2, 3 mM phenylalanine precursor on the accumulation of polyphenolics was studied.

4. Evaluating the calli extracts from chemical and biological points of view and comparing it to that of the locally cultivated plant extracts:

All elicited (treated) calli hydro-alcoholic extracts were compared with extracts from untreated callus and both were compared with the cultivated intact plant. Total polyphenolic content was measured using follin-Ciocalteau method, followed by caffeic acids derivatives analysis and quantitation using LC/MS/MS.

The highest total polyphenolic content was achieved and detected in hydro-alcoholic extracts of calli treated with **10 mg/L MeJA**, with a statistically significant **1.81-fold** increase as compared to the cultivated plant, followed by calli treated with **2 mM Phenylalanine** that showed a statistically significant **1.5-fold** increase in polyphenolic content in comparison with the cultivated plant. There was no significant increase was observed in caffeic acid derivatives yield in the elicited calli extracts. Also, the efficacy of the extracts of treated and untreated calli were compared with the cultivated plant extracts, all *Echinacea* extracts were examined for their cytotoxicity, anti-oxidant activity, immunostimulatory activity, enhancement of macrophage and neutrophil phagocytic activity and in-vitro anti-inflammatory activity. It was observed that the hydro-alcoholic extracts of calli treated with **10 mg/L MeJA**, **2 mM Phe** and **2 μ M CuSO₄** exhibited the most potent activities among all the tested extracts.

9. APPENDIX

All Experimental results were presented as Mean \pm SEM of three parallel measurements. Statistical evaluation was carried out by one-way analysis of variance (ANOVA). Statistical significance is expressed as $p < 0.05$ where,

a: Effective concentration value is statistically significant compared to its value for Callus extract

b: Effective concentration value is statistically significant compared to its value for Cultivated extract

c: Effective concentration value is statistically significant compared to its value for Phe 2 extract

d: Effective concentration value is statistically significant compared to its value for MeJA 10 extract

e: Effective concentration value is statistically significant compared to its value for SNP 50 extract

f: Effective concentration value is statistically significant compared to its value for MeJA 5 extract

g: Effective concentration value is statistically significant compared to its value for EtOH 0.5 extract

h: Effective concentration value is statistically significant compared to its value for CuSO₄ 2 extract

i: Effective concentration value is statistically significant compared to its value for EtOH 1 extract

j: Effective concentration value is statistically significant compared to its value for Control acetic acid extract

k: Effective concentration value is statistically significant compared to its value for CuSO₄ 4 extract

l: Effective concentration value is statistically significant compared to its value for Chitosan 5 extract

m: Effective concentration value is statistically significant compared to its value for Chitosan 50 extract

n: Effective concentration value is statistically significant compared to its value for SNP 250 extract

o: Effective concentration value is statistically significant compared to its value for Phe 1 extract

p: Effective concentration value is statistically significant compared to its value for MeJA 20 extract

q: Effective concentration value is statistically significant compared to its value for Phe 3 extract

r: Effective concentration value is statistically significant compared to its value for SNP 100 extract

s: Effective concentration value is statistically significant compared to its value for Chitosan 25 extract

t: Effective concentration value is statistically significant compared to its value for EtOH 2 extract

u: Effective concentration value is statistically significant compared to its value for CuSO₄ 8 extract

Table 4: Total polyphenolic content results

Extract Identity	Concentration in ($\mu\text{g GAE}/2 \text{ mg DW}$)
Callus	21.4200 \pm .40 c,d,e,f,g,h,i,j,k,l,m,o,p,q,r,s,t,u
Cultivated	22.0200 \pm .00 c,d,e,f,g,h,i,j,k,l,m,o,p,q,r,s,t,u
Phe 2	33.2200 \pm .00 a,b,d,e,f,g,h,i,j,k,l,m,n,o,p,q,r,s,t,u
MeJA 10	40.0200 \pm .00 a,b,c,e,f,g,h,i,j,k,l,m,n,o,p,q,r,s,t,u
SNP 50	9.7200 \pm .900 a,b,c,d,h,n,o,p,s,u
MeJA 5	12.1200 \pm 1.70 a,b,c,d,g,h,i,j,k,l,n,o,p,q,s,u
EtOH 0.5	8.8200 \pm .6000 a,b,c,d,f,h,m,n,o,p,s,t,u
CuSO ₄ 2	17.6200 \pm .400 a,b,c,d,e,f,g,i,j,k,l,m,n,o,p,q,r,t
EtOH 1	9.6200 \pm .400 a,b,c,d,f,h,n,o,p,s,u
Control acetic acid	9.5200 \pm .500 a,b,c,d,f,h,n,o,p,s,u
CuSO ₄ 4	8.1200 \pm 1.10 a,b,c,d,f,h,m,n,o,p,r,s,t,u
Chitosan 5	9.5200 \pm .7000 a,b,c,d,f,h,n,o,p,s,u
Chitosan 50	11.4200 \pm .400 a,b,c,d,g,h,k,n,o,p,q,s,u
SNP 250	21.9200 \pm .900 c,d,e,f,g,h,i,j,k,l,m,o,p,q,r,s,t,u
Phe 1	4.3200 \pm .1000 a,b,c,d,e,f,g,h,i,j,k,l,m,n,q,r,s,t,u
MeJA 20	4.3200 \pm .1000 a,b,c,d,e,f,g,h,i,j,k,l,m,n,q,r,s,t,u
Phe 3	8.1200 \pm .3000 a,b,c,d,f,h,m,n,o,p,r,s,t,u
SNP 100	11.1200 \pm .90 a,b,c,d,h,k,n,o,p,q,s,u
Chitosan 25	18.8200 \pm .800 a,b,c,d,e,f,g,i,j,k,l,m,n,o,p,q,r,t
EtOH 2	11.5200 \pm .100 a,b,c,d,f,g,h,k,n,o,p,q,s,u
CuSO ₄ 8	17.4200 \pm 2.4 a,b,c,d,e,f,g,i,j,k,l,m,n,o,p,q,r,t

Table 5: Total antioxidant activity

Extract Identity	IC50 values (µg/ml) of ABTS scavenging activity
Callus	39.6367±1.06529 c,f,i,j,k,m,o,t,u
Cultivated	30.8748±1.10652 f,i,j,k,m,o,t,u
Phe 2	7.1797±1.39872 a,e,f,i,j,k,l,m,o,q,t,u
MeJA 10	26.6113±.01798 f,i,j,k,m,o,q,t,u
SNP 50	47.6856±3.03857 c,f,g,i,j,m,o,s,t,u
MeJA 5	77.3569±1.87402 a,b,c,d,e,g,h,i,j,l,m,n,o,p,r,s,t,u
EtOH 0.5	16.7225±2.27150 e,f,i,j,k,m,o,q,t,u
CuSO ₄ 2	18.8335±2.11050 f,i,j,k,m,o,q,t,u
EtOH 1	1.1501E2±1.88550 a,b,c,d,e,f,g,h,j,k,l,m,n,o,p,q,r,s,u
Control acetic acid	1.6665E2±1.66150 a,b,c,d,e,f,g,h,i,k,l,m,n,o,p,q,r,s,t,u
CuSO ₄ 4	72.2770±1.89400 a,b,c,d,e,f,g,h,i,j,m,n,o,p,q,r,s,t,u
Chitosan 5	45.055±5.83250 c,f,i,j,m,o,t,u
Chitosan 50	4.8301E2±19.45350 a,b,c,d,e,f,g,h,i,j,k,l,n,o,p,q,r,s,t,u
SNP 250	23.6320±.02800 f,i,j,k,m,o,q,t,u
Phe 1	2.0989E2±1.11750 a,b,c,d,e,f,g,h,i,j,k,l,m,n,p,q,r,s,t
MeJA 20	27.4600±1.37501 f,i,j,k,m,o,q,t,u
Phe 3	57.9615±1.62950 c,d,g,h,i,j,m,n,o,p,r,s,t,u
SNP 100	19.4496±.05920 f,i,j,k,m,o,q,t,u
Chitosan 25	15.9717±3.88189 e,f,i,j,k,m,o,q,t,u
EtOH 2	1.2182E2±1.09152 a,b,c,d,e,f,g,h,j,k,l,m,n,o,p,q,r,s,u
CuSO ₄ 8	2.0805E2±40.68719 a,b,c,d,e,f,g,h,i,j,k,l,m,n,p,q,r,s,t

Table 6: *In vitro* phagocytic activity assay using Polymorphonuclear neutrophils

Extract Identity	EC₅₀ values (µg/ml) of neutrophil phagocytosis activity	EC₅₀ value (µg/ml) of neutrophil yeast killing
Callus	2.0131E2±5.1315 b,c,d,e,f,g,h,i,j,k,l,m,n,o,p,q,r,s,t,u	1.1112E2±1.30550 e,g,j,k,m,n,q,r,s
Cultivated	40.9805±1.46650 a,c,d,e,g,h,i,j,k,l,m,n,o,p,t,u	56.2915±2.05450 e,j,m,n,q,s
Phe 2	7.7215±.31450 a,b,e,f,i,j,k,l,m,n,o,p,q,r,s,t,u	50.2625±.04650 e,j,m,n,q,s
MeJA 10	6.2045±.1155 a,b,e,f,i,j,k,l,m,n,o,p,q,r,s,t,u	22.8230±1.34000 e,f,j,m,n,q,s
SNP 50	85.8410±5.49400 a,b,c,d,f,g,h,i,j,k,l,m,n,q,r,s,u	3.0549E2±2.12450 a,b,c,d,f,g,h,i,j,k,l,m,n,o,p,q,r,s,t,u
MeJA 5	48.5935±2.50050 a,c,d,e,g,h,i,j,k,l,m,n,o,p,r,t,u	1.4006E2±1.95950 d,e,g,h,j,k,m,n,p,q,r,s,t
EtOH 0.5	1.0985±.0525 a,b,e,f,i,j,k,l,m,n,o,p,q,r,s,t,u	3.2750±.27200 a,e,f,i,j,l,m,n,o,q,s
CuSO ₄ 2	4.9390±.7850 a,b,e,f,i,j,k,l,m,n,o,p,q,r,s,t,u	33.7615±.08150 e,f,i,j,m,n,q,s
EtOH 1	3.0596E2±5.8315 a,b,c,d,e,f,g,h,j,k,l,m,n,o,p,q,r,s,t	1.3991E2±4.25050 d,e,g,h,j,k,m,n,p,q,r,s,t
Control acetic acid	1.7534E2±2.9100 a,b,c,d,e,f,g,h,i,k,l,m,n,o,p,q,r,s,t,u	1.4820E3±1.39218E2 a,b,c,d,e,f,g,h,i,k,l,m,n,o,p,q,r,s,t,u
CuSO ₄ 4	1.2662E3±.0000 a,b,c,d,e,f,g,h,i,j,l,m,n,o,p,q,r,s,t,u	6.9285±2.07350 a,e,f,i,j,l,m,n,o,q,s
Chitosan 5	7.6259E2±26.635 a,b,c,d,e,f,g,h,i,j,k,m,n,o,p,q,r,s,t,u	1.0254E2±8.14700 e,g,j,k,m,n,q,r,s
Chitosan 50	3.5878E2±.13200 a,b,c,d,e,f,g,h,i,j,k,l,n,o,p,q,r,s,t,u	5.0473E2±13.21000 a,b,c,d,e,f,g,h,l,j,k,l,n,o,p,q,r,s,t,u
SNP 250	3.3584E2±1.6965 a,b,c,d,e,f,g,h,i,j,k,l,m,o,p,q,r,s,t,u	1.2933E3±15.55250 a,b,c,d,e,f,g,h,i,j,k,l,m,o,p,q,r,s,t,u
Phe 1	68.8815±1.99350 a,b,c,d,f,g,h,i,j,k,l,m,n,r,u	1.0554E2±5.66050 e,g,j,k,m,n,q,r,s
MeJA 20	70.8180±.33000 a,b,c,d,f,g,h,i,j,k,l,m,n,q,r,u	32.4735±.06050 e,f,i,j,m,n,q,s
Phe 3	50.8900±.93600 a,c,d,e,g,h,i,j,k,l,m,n,p,r,t,u	6.5797E2±4.99500 a,b,c,d,e,f,g,h,i,j,k,l,m,n,o,p,r,t,u
SNP 100	27.2250±2.03500 a,c,d,e,f,g,h,i,j,k,l,m,n,o,p,q,s,t0,u	1.8505±.00150 a,e,f,i,j,l,m,n,o,q,s
Chitosan 25	52.0435±2.66250 a,c,d,e,g,h,i,j,k,l,m,n,r,t,u	7.1812E2±.99300 a,b,c,d,e,f,g,h,i,j,k,l,m,n,o,p,r,t,u
EtOH 2	75.5715±2.86350 a,b,c,d,f,g,h,i,j,k,l,m,n,q,r,s,u	35.8020±4.38000 e,f,i,j,m,n,q,s
CuSO ₄ 8	3.1603E2±4.2850 a,b,c,d,e,f,g,h,j,k,l,m,n,o,p,q,r,s,t	80.2885±2.8945 e,j,m,n,q,s

Table 7: *In vitro* phagocytic activity assay using peritoneal macrophages

Extract Identity	Macrophage phagocytosis	Macrophage killing
Callus	1.5526E5±4.20072E2 d,e,k,l,q	1.4331E2±4.31800 e,i,p
Cultivated	5.5813E4±3.76028E3 d,e,k,l,q	37.9610±2.33100 e,i,p
Phe 2	5.4885E4±3.00251E3 d,e,k,l,q	58.9275±2.80250 e,i,p
MeJA 10	4.8736E5±3.97574E5 a,b,c,g,h,j,k,l,m,n,o,p,r,s,t,u	58.7630±5.48600 e,i,p
SNP 50	4.6772E5±7.29453E3 a,b,c,g,h,k,l,m,n,o,p,q,r,s,t,u	4.6325E3±2.23653E2 a,b,c,d,f,g,h,I,j,k,l,m,n,o,p,q,r,s,t,u
MeJA 5	2.2444E5±1.82088E5 k,l,q	70.1625±.96550 e,i,p
EtOH 0.5	2.7048E3±8.4645 d,e,k,l,q	2.1475±.19650 e,i,j,p
CuSO ₄ 2	7.2694E3±28.99450 d,e,k,l,q	16.1375±1.64950 e,i,j,p
EtOH 1	2.6454E5±3.98021E3 k,l,q	1.4741E3±1.42694E2 a,b,c,d,e,f,g,h,j,k,l,m,n,o,p,q,r,s,t,u
Control acetic acid	1.9051E5±6.10151E3 d,k,lq	1.9845E2±12.8725 e,g,h,i,q,r,s,t
CuSO ₄ 4	2.0523E6±3.92084E4 a,b,c,d,e,f,g,h,i,j,l,m,n,o,p,q,r,s,t,u	1.0148E2±2.23450 e,i,p
Chitosan 5	9.3297E5±8.32530E3 a,b,c,d,e,f,g,h,i,j,k,m,n,o,p,r,s,t,u	1.5555E2±8.04800 e,i,p
Chitosan 50	1.6014E5±9.08919E3 d,e,k,l,q	1.0554E2±3.71850 e,i,p
SNP 250	7.2168E4±3.27062E3 d,e,k,l,q	1.5612E2±.27150 e,i,p
Phe 1	6.1102E4±5.02436E3 d,e,k,l,q	1.3009E2±9.08700 e,i,p
MeJA 20	1.6900E5±1.72142E4 d,e,k,l,q	3.3915E2±23.0725 a,b,c,d,e,f,g,h,I,k,l,m,n,o,q,r,s,t,u
Phe 3	7.5254E5±1.52674E4 a,b,c,e,f,g,h,I,j,k,m,n,o,p,r,s,t,u	6.0430±.29700 e,i,j,p
SNP 100	1.1170E4±2.70671E2 d,e,k,l,q	4.7955±.12950 e,i,j,p
Chitosan 25	5.2126E4±2.33536E2 d,e,k,l,q	7.1830±.11900 e,i,j,p
EtOH 2	6.4927E4±1.63950E3 d,e,k,l,q	5.9160±.49100 e,i,j,p
CuSO ₄ 8	1.5357E5±4.99336E3 k,l,q	37.3645±1.55050 e,i,p

Table 8: *In-vitro* immunostimulatory Activity

Extract Identity	EC₅₀ values (expressed in in µg/ml) causing a lymphocyte stimulation index of 3
Callus	1.5865E3±1.44479E2 b,c,d,e,f,g,h,i,j,k,l,m,n,p,q,r,s,t,u
Cultivated	1.6366E2±8.67650 a,c,e,i,j,k,l,n,o,q,s,u
Phe 2	2.5439E2±19.31100 a,d,e,g,i,j,k,l,n,o,p,q,s,t,u
MeJA 10	1.3332E2±.15900 a,c,e,i,j,k,l,m,n,o,q,s,u
SNP 50	5.3718E2±45.80950 a,b,c,d,f,g,h,j,k,l,m,n,o,p,q,r,t,u
MeJA 5	1.8513E2±5.04800 a,e,g,l,j,k,l,n,o,q,s,t,u
EtOH 0.5	72.5350±2.24300 a,c,e,f,g,h,i,j,k,l,m,n,o,q,s,u
CuSO ₄ 2	2.1224E2±24.92800 a,e,g,i,j,k,l,n,o,q,s,t,u
EtOH 1	5.6548E2±19.65100 a,b,c,d,f,g,h,j,k,l,m,n,o,p,q,r,s,t,u
Control acetic acid	4.1465E2±1.71600 a,b,c,d,e,f,g,h,i,k,m,n,o,p,r,t,u
CuSO ₄ 4	4.7508E3±.00000 a,b,c,d,e,f,g,h,i,j,l,m,n,o,p,q,r,s,t,u
Chitosan 5	4.0998E2±3.31750 a,b,c,d,e,f,g,h,i,k,l,m,n,o,p,r,t,u
Chitosan 50	2.6307E2±6.02400 a,d,e,g,i,j,k,l,m,n,o,p,q,s,t,u
SNP 250	1.9565E3±29.60300 a,b,c,d,e,f,g,h,i,j,k,l,m,o,p,q,r,s,t,u
Phe 1	1.5191E3±28.98850 b,c,d,e,f,g,h,i,j,k,l,m,n,p,q,r,s,t,u
MeJA 20	1.3752E2±11.78350 a,c,e,i,j,k,l,m,n,o,q,s,u
Phe 3	3.8066E2±3.55250 a,b,c,d,e,f,g,h,i,k,m,n,o,p,r,t,u
SNP 100	1.6861E2±4.41050 a,c,e,i,j,k,l,n,o,q,s,u
Chitosan 25	4.3339E2±31.08000 a,b,c,d,e,f,g,h,i,k,m,n,o,p,r,t,u
EtOH 2	74.1580±2.36600 a,c,e,f,h,l,j,k,l,m,n,o,q,s,u
CuSO ₄ 8	3.2551E3±34.89100 a,b,c,d,e,f,g,h,i,j,k,l,m,n,o,p,q,r,s,t

Table 9: *In vitro* anti-inflammatory activity

Extract Identity	Effective concentration in µg/ml causing a lymphocyte stimulation index of 3
Callus	1.5903E2±6.70950 b,c,d,e,f,g,h,i,j,k,l,m,n,o,p,q,r,s,t,u
Cultivated	18.0095±.13850 a,d,e,g,h,i,j,k,l,m,n,q,r,s,t,u
Phe 2	18.3355±.09050 a,d,e,f,g,h,i,j,k,l,m,n,q,r,s,t,u
MeJA 10	6.3525±.21450 a,b,c,e,f,i,j,k,l,m,o,p,u
SNP 50	46.0320±1.53800 a,b,c,d,f,g,h,i,j,k,l,m,n,o,p,q,r,s,t,u
MeJA 5	12.8005±1.74150 a,d,e,g,h,i,j,k,l,m,n,o,p,q,r,t,u
EtOH 0.5	3.8630±.00300 a,b,c,e,f,i,j,k,l,m,o,p,u
CuSO ₄ 2	1.3525±.31450 a,b,c,e,f,i,j,k,l,m,o,p,u
EtOH 1	60.4690±.57000 a,b,c,d,e,f,g,h,k,l,m,n,o,p,q,r,s,t,u
Control acetic acid	64.8825±1.36750 a,b,c,d,e,f,g,h,k,l,m,n,o,p,q,r,s,t,u
CuSO ₄ 4	79.5135±2.01350 a,b,c,d,e,f,g,h,i,j,l,m,n,o,p,q,r,s,t,u
Chitosan 5	92.7165±2.85350 a,b,c,d,e,f,g,h,i,j,k,m,n,o,p,q,r,s,t
Chitosan 50	1.0072E2±3.14150 a,b,c,d,e,f,g,h,i,j,k,l,n,o,p,q,r,s,t
SNP 250	2.0270±.42000 a,b,c,e,f,i,j,k,l,m,o,p,u
Phe 1	22.4015±2.23650 a,d,e,f,g,h,i,j,k,l,m,n,q,r,s,t,u
MeJA 20	20.3855±1.37350 a,d,e,f,g,h,i,j,k,l,m,n,q,r,s,t,u
Phe 3	5.1790±.85500 a,b,c,e,f,i,j,k,l,m,o,p,u
SNP 100	4.4785±.29650 a,b,c,e,f,i,j,k,l,m,o,p,u
Chitosan 25	7.3335±.59850 a,b,c,e,i,j,k,l,m,o,p,u
EtOH 2	4.1375±.01250 a,b,c,e,f,i,j,k,l,m,o,p,u
CuSO ₄ 8	97.5910±3.03600 a,b,c,d,e,f,g,h,i,j,k,n,o,p,q,r,s,t

Table 10: Nitric oxide scavenging activity

Extract Identity	EC50 in µg/ml
Callus	3.3939E2±15.81800 b,c,d,e,f,g,h,j,l,m,n,o,p,q,r,s,u
Cultivated	1.3030E2±.241 a,d,e,g,h,i,k,l,m,n,o,q,s,t
Phe 2	28.9375±.0175 a,b,e,i,k,l,m,n,o,q,t
MeJA 10	6.3730±.49800 a,b,e,i,k,l,m,n,o,q,t
SNP 50	5.3849E2±2.55200 a,b,c,d,f,g,h,i,j,k,l,m,n,o,p,q,r,s,t,u
MeJA 5	51.7450±2.66800 a,e,i,k,l,m,n,o,q,t
EtOH 0.5	5.2175±.08050 a,b,e,i,k,l,m,n,o,q,t
CuSO ₄ 2	22.4880±2.882 a,b,e,i,k,l,m,n,o,q,t
EtOH 1	2.9588E2±6.96750 b,c,d,e,f,g,h,j,l,m,n,o,p,q,r,s,u
Control acetic acid	61.6670±8.180 a,e,i,k,l,m,n,o,q,t
CuSO ₄ 4	2.8318E2±14.4275 b,c,d,e,f,g,h,j,l,m,n,o,p,q,r,s,u
Chitosan 5	7.7059E2±41.86150 a,b,c,d,e,f,g,h,i,j,k,m,n,p,q,r,s,t,u
Chitosan 50	1.1503E3±1.05772E2 a,b,c,d,e,f,g,h,i,j,k,l,n,o,p,q,r,s,t,u
SNP 250	1.3146E3±56.44400 a,b,c,d,e,f,g,h,i,j,k,l,m,o,p,q,r,s,t,u
Phe 1	8.3263E2±18.92000 a,b,c,d,e,f,g,h,i,j,k,m,n,p,q,r,s,t,u
MeJA 20	68.373±3.53750 a,e,i,k,l,m,n,o,q,t
Phe 3	6.7502E2±000 a,b,c,d,e,f,g,h,i,j,k,l,m,n,o,p,r,s,t,u
SNP 100	87.8045±3.93250 a,e,i,k,l,m,n,o,q,t
Chitosan 25	32.7320±.09800 a,b,e,i,k,l,m,n,o,q,t
EtOH 2	3.2755E2±3.30100 b,c,d,e,f,g,h,j,l,m,n,o,p,q,r,s,u
CuSO ₄ 8	49.9875±8.25650 a,b,e,i,k,l,m,n,o,q,t

10. REFERENCES

- 1 Perry, N. B., Burgess, E. J. & Glennie, V. L. Echinacea standardization: analytical methods for phenolic compounds and typical levels in medicinal species. *Journal of Agricultural and Food Chemistry* **49**, 1702-1706 (2001).
- 2 Abbasi, B. H., Saxena, P. K., Murch, S. J. & Liu, C. Echinacea biotechnology: challenges and opportunities. *In Vitro Cellular & Developmental Biology-Plant* **43**, 481-492 (2007).
- 3 Bałan, B., Róewski, F., Zdanowski, R. & Skopińska-Róewska, E. Immunotropic activity of Echinacea. Part I. History and chemical structure. *Central European Journal of Immunology* **37**, 46 (2012).
- 4 Mckeown, K. A. in *Perspectives on new crops and new uses*, A review of the taxonomy of the genus Echinacea 482-489 (ASHS Press, Alexandria, Va, 1999).
- 5 Qu, L., Chen, Y., Wang, X., Scalzo, R. & Davis, J. M. Patterns of variation in alkamides and cichoric acid in roots and aboveground parts of Echinacea purpurea (L.) Moench. *HortScience* **40**, 1239-1242 (2005).
- 6 Barrett, B. Medicinal properties of Echinacea: A critical review. *Phytomedicine* **10**, 66-86 (2003).
- 7 Foster, S. in *Special forest products: Biodiversity Meets the Marketplace*, Medicinal plant development in the United States (Washington DC, 1997).
- 8 Senchina, D. S. *Medical botany of the genus Echinacea* Doctor of Philosophy thesis, Iowa State University, (2006).
- 9 Gupta, M., Sharma, D., Sharma, A., Kumari, V. & Goshain, O. A Review on Purple Cone Flower (Echinacea purpurea L. Moench). *Journal of Pharmacy Research* **5** (2012).
- 10 Upton, R., Graff, A., Jolliffe, G., Länger, R. & Williamson, E. *American Herbal Pharmacopoeia: botanical pharmacognosy-microscopic characterization of botanical medicines*. (CRC Press Inc., 2011).
- 11 Galambosi, B. in *Echinacea*, 4 Cultivation in Europe 29 (CRC Press, 2004).
- 12 Letchamo, W., Polydeonny, L., Gladisheva, N., Arnason, T., Livesey, J. & Awang, D. in *Trends in new crops and new uses*, Factors affecting Echinacea quality 514-521 (ASHS Press, Alexandria, Va, 2002).
- 13 El-Gengaihi, S., Shalaby, A., Agina, E. & Hendawy, S. Alkylamides of Echinacea purpurea L. as influenced by plant ontogony and fertilization. *Journal of herbs, spices & medicinal plants* **5**, 35-41 (1998).
- 14 Sf, H. *Agriculture and Chemical Studies on Echinacea purpurea* Master Degree thesis, Zagazig University, (1995).
- 15 Shalaby, A., Agina, E., El-Gengaihi, S., El-Khayat, A. & Hindawy, S. Response of Echinacea to some agricultural practices. *Journal of herbs, spices & medicinal plants* **4**, 59-67 (1997).
- 16 El-Sayed, A., Shalaby, A., El-Hanafy, H. & El-Razik, T. A. Effects of Chemical Fertilizers on Growth and Active Constituents of Echinacea paradoxa L. Plants. *Journal of Horticultural Science & Ornamental Plants* **4**, 125-133 (2012).
- 17 Orhan, I. *Biotechnological Production of Plant Secondary Metabolites*. (Bentham Science Publishers, 2012).
- 18 Bhojwani, S. S. & Dantu, P. K. in *Plant Tissue Culture: An Introductory Text*, Production of Industrial Phytochemicals 275-286 (Springer, 2013).

- 19 Coker, P. S., Camper, N. D. & Miller, S. in *Echinacea: The genus Echinacea*, In vitro culture of Echinacea species 23-28 (CRC Press, 2004).
- 20 Murthy, H. N., Hahn, E. & Paek, K. Adventitious roots and secondary metabolism. *Chinese Journal of Biotechnology* **24**, 711-716 (2008).
- 21 Wu, C., Dewir, Y. H., Hahn, E. & Paek, K. Optimization of culturing conditions for the production of biomass and phenolics from adventitious roots of Echinacea angustifolia. *Journal of Plant Biology* **49**, 193-199 (2006).
- 22 Wu, C., Murthy, H. N., Hahn, E. & Paek, K. Large-scale cultivation of adventitious roots of Echinacea purpurea in airlift bioreactors for the production of chichoric acid, chlorogenic acid and caftaric acid. *Biotechnology letters* **29**, 1179-1182 (2007).
- 23 Hahn, E., Wu, C. & Paek, K. in *VI International Symposium on In Vitro Culture and Horticultural Breeding* 829. 73-78 (2009).
- 24 Jeong, J., Wu, C., Murthy, H. N., Hahn, E. & Paek, K. Application of an airlift bioreactor system for the production of adventitious root biomass and caffeic acid derivatives of Echinacea purpurea. *Biotechnology and Bioprocess Engineering* **14**, 91-98 (2009).
- 25 Namdeo, A. Plant cell elicitation for production of secondary metabolites: a review. *Pharmacognosy reviews* **1**, 69-79 (2007).
- 26 Ruiz-García, Y. & Gómez-Plaza, E. Elicitors: A tool for improving fruit phenolic content. *Agriculture* **3**, 33-52 (2013).
- 27 Cai, Z., Kastell, A., Mewis, I., Knorr, D. & Smetanska, I. Polysaccharide elicitors enhance anthocyanin and phenolic acid accumulation in cell suspension cultures of Vitis vinifera. *Plant Cell, Tissue and Organ Culture (PCTOC)* **108**, 401-409 (2012).
- 28 Kim, H., Chen, F., Wang, X. & Rajapakse, N. C. Effect of methyl jasmonate on secondary metabolites of sweet basil (Ocimum basilicum L.). *Journal of agricultural and food chemistry* **54**, 2327-2332 (2006).
- 29 Li, W. & Barz, W. Structure and Accumulation of Phenolics in Elicited Echinacea purpurea cell cultures. *Planta medica* **72**, 248-254 (2006).
- 30 Shohael, A. M., Murthy, H. N., Hahn, E. & Paek, K. Methyl jasmonate induced overproduction of eleutherosides in somatic embryos of Eleutherococcus senticosus cultured in bioreactors. *Electronic Journal of Biotechnology* **10**, 633-637 (2007).
- 31 Krzyzanowska, J., Czubacka, A., Pecio, L., Przybys, M., Doroszewska, T., Stochmal, A. & Oleszek, W. The effects of jasmonic acid and methyl jasmonate on rosmarinic acid production in Mentha piperita cell suspension cultures. *Plant Cell, Tissue and Organ Culture (PCTOC)* **108**, 73-81 (2012).
- 32 Mobin, M., Wu, C., Tewari, R. K. & Paek, K. Studies on the glyphosate-induced amino acid starvation and addition of precursors on caffeic acid accumulation and profiles in adventitious roots of Echinacea purpurea (L.) Moench. *Plant Cell, Tissue and Organ Culture (PCTOC)* **120**, 291-301 (2015).
- 33 El-Naggar, H. M. Phenylalanine ammonia-lyase (PAL) gene activity in response to proline and tyrosine in rosemary callus culture. *African Journal of Biotechnology* **11**, 159-163 (2013).
- 34 Roy, D. & Mukhopadhyay, S. Enhanced rosmarinic acid production in cultured plants of two species of Mentha. *Indian journal of experimental biology* **50**, 817-825 (2012).
- 35 Liu, C., Abbasi, B. H., Gao, M., Murch, S. J. & Saxena, P. K. Caffeic acid derivatives production by hairy root cultures of Echinacea purpurea. *Journal of agricultural and food chemistry* **54**, 8456-8460 (2006).

- 36 Wang, B., Zhang, G., Zhu, L., Chen, L. & Zhang, Y. Genetic transformation of Echinacea purpurea with Agrobacterium rhizogenes and bioactive ingredient analysis in transformed cultures. *Colloids and Surfaces B: Biointerfaces* **53**, 101-104 (2006).
- 37 Abbasi, B. H., Tian, C.-L., Murch, S. J., Saxena, P. K. & Liu, C.-Z. Light-enhanced caffeic acid derivatives biosynthesis in hairy root cultures of Echinacea purpurea. *Plant cell reports* **26**, 1367-1372 (2007).
- 38 Romero, F. R., Delate, K., Kraus, G. A., Solco, A. K., Murphy, P. A. & Hannapel, D. J. Alkamide production from hairy root cultures of Echinacea. *In Vitro Cellular & Developmental Biology-Plant* **45**, 599-609 (2009).
- 39 Liu, R., Li, W., Sun, L. & Liu, C. Improving root growth and cichoric acid derivatives production in hairy root culture of Echinacea purpurea by ultrasound treatment. *Biochemical Engineering Journal* **60**, 62-66 (2012).
- 40 Bohlmann, F. & Hoffmann, H. Further amides from Echinacea purpurea. *Phytochemistry* **22**, 1173-1175 (1983).
- 41 Bae, J. *Synthesis of natural compounds in echinacea*. (Iowa State University, 2006).
- 42 Bauer, V., Jurcic, K., Puhlmann, J. & Wagner, H. Immunologic in vivo and in vitro studies on Echinacea extracts. *Arzneimittel-forschung* **38**, 276-281 (1988).
- 43 Bauer, R., Remiger, P. & Wagner, H. Alkamides from the roots of Echinacea angustifolia. *Phytochemistry* **28**, 505-508 (1989).
- 44 Upton, R. *Echinacea Purpurea Root: Echinacea Purpurea (L.) Moench: Standards of Analysis, Quality Control, and Therapeutics*. (American Herbal Pharmacopoeia, 2004).
- 45 Clifford, L. J. *Isolation and Characterization of Anti-oxidant and Anti-inflammatory Constituents from Echinacea Purpurea (L.) Moench*, Michigan State University. Department of Food Science and Human Nutrition, (2002).
- 46 Lim, T. K. *Edible medicinal and non-medicinal plants*. Vol. 1 (Springer, 2012).
- 47 Bone, K. & Mills, S. *Principles and Practice of Phytotherapy: Modern Herbal Medicine*. (Elsevier Health Sciences, 2013).
- 48 Bałan, B. J., Rózewski, F., Skopińska-Rózewska, E., Wojdas, A., Zdanowski, R. & Stankiewicz, W. Immunotropic activity of Echinacea. Part II. Experimental and clinical data. *Central European Journal of Immunology* **37**, 52 (2012).
- 49 Bauer, R. & Wagner, H. *Echinacea species as potential immunostimulatory drugs*. Vol. 5 253-321 (Economic and medicinal plant research, 1991).
- 50 Bisset, N. *Herbal drugs and phytopharmaceuticals*. (London: CRC Press, 1994).
- 51 Bodinet, C., Willigmann, I. & Beuscher, N. Host-resistance increasing activity of root extracts from Echinacea species. *Planta Medica (Germany)* **59**, A672-673 (1993).
- 52 Stotzem, C., Hungerland, U. & Mengs, U. Influence of Echinacea purpurea on the phagocytosis of human granulocytes. *Medical science research* **20**, 719-720 (1992).
- 53 Currier, N. & Miller, S. Natural killer cells from aging mice treated with extracts from Echinacea purpurea are quantitatively and functionally rejuvenated. *Experimental gerontology* **35**, 627-639 (2000).
- 54 Sun, L. Z., Currier, N. L. & Miller, S. C. The American coneflower: a prophylactic role involving nonspecific immunity. *The Journal of Alternative and Complementary Medicine* **5**, 437-446 (1999).
- 55 Burger, R. A., Torres, A. R., Warren, R. P., Caldwell, V. D. & Hughes, B. G. Echinacea-induced cytokine production by human macrophages. *International journal of immunopharmacology* **19**, 371-379 (1997).

- 56 Melchart, D., Linde, K., Worku, F., Sarkady, L., Holzmann, M., Jurcic, K. & Wagner, H. Results of five randomized studies on the immunomodulatory activity of preparations of Echinacea. *The Journal of Alternative and Complementary Medicine* **1**, 145-160 (1995).
- 57 Morazzoni, P., Cristoni, A., Di Pierro, F., Avanzini, C., Ravarino, D., Stornello, S., Zucca, M. & Musso, T. In vitro and in vivo immune stimulating effects of a new standardized Echinacea angustifolia root extract (Polinacea™). *Fitoterapia* **76**, 401-411 (2005).
- 58 Percival, S. S. Use of Echinacea in medicine. *Biochemical pharmacology* **60**, 155-158 (2000).
- 59 Roesler, J., Emmendorffer, A., Steinmüller, C., Luettig, B., Wagner, H. & Lohmann-Matthes, M. Application of purified polysaccharides from cell cultures of the plant Echinacea purpurea to test subjects mediates activation of the phagocyte system. *International journal of immunopharmacology* **13**, 931-941 (1991).
- 60 Schwarz, E., Metzler, J., Diedrich, J. P., Freudenstein, J., Bode, C. & Bode, J. C. Oral administration of freshly expressed juice of Echinacea purpurea herbs fail to stimulate the nonspecific immune response in healthy young men: results of a double-blind, placebo-controlled crossover study. *Journal of Immunotherapy* **25**, 413-420 (2002).
- 61 See, D. M., Broumand, N., Sahl, L. & Tilles, J. G. In vitro effects of echinacea and ginseng on natural killer and antibody-dependent cell cytotoxicity in healthy subjects and chronic fatigue syndrome or acquired immunodeficiency syndrome patients. *Immunopharmacology* **35**, 229-235 (1997).
- 62 Mistrikova, I. & Vaverkova, S. Echinacea—chemical composition, immunostimulatory activities and uses. *Thaizia J Bot* **16**, 11-26 (2006).
- 63 Mazzi, E. A. & Soliman, K. F. In vitro screening for the tumoricidal properties of international medicinal herbs. *Phytotherapy research* **23**, 385-398 (2009).
- 64 Rogala, E., Skopinska-Rozewska, E., Wasiutynski, A., Siwicki, A. K., Sommer, E. & Pastewka, K. Echinacea purpurea diminishes neovascular reaction induced in mice skin by human cancer cells and stimulates non-specific cellular immunity in humans. *Central European Journal of Immunology* **33**, 127 (2008).
- 65 Skaudickas, D., Kondrotas, A., Kevelaitis, E. & Venskutonis, P. The effect of Echinacea purpurea (L.) Moench extract on experimental prostate hyperplasia. *Phytotherapy Research* **23**, 1474-1478 (2009).
- 66 Chicca, A., Adinolfi, B., Martinotti, E., Fogli, S., Breschi, M., Pellati, F., Benvenuti, S. & Nieri, P. Cytotoxic effects of Echinacea root hexanic extracts on human cancer cell lines. *Journal of ethnopharmacology* **110**, 148-153 (2007).
- 67 Tsai, Y., Chiou, S., Chan, K., Sung, J. & Lin, S. Caffeic acid derivatives, total phenols, antioxidant and antimutagenic activities of Echinacea purpurea flower extracts. *LWT-Food Science and Technology* **46**, 169-176 (2012).
- 68 Facino, R. M., Carini, M., Aldini, G., Marinello, C., Arlandini, E., Franzoi, L., Colombo, M., Pietta, P. & Mauri, P. Direct characterization of caffeoyl esters with antihyaluronidase activity in crude extracts from Echinacea angustifolia roots by fast atom bombardment tandem mass spectrometry. *Farmaco (Societa chimica italiana: 1989)* **48**, 1447-1461 (1993).
- 69 Facino, R. M., Carini, M., Aldini, G., Saibene, L., Pietta, P. & Mauri, P. Echinacoside and Caffeoyl Conjugates Protect Collagen from Free Radical-Induced Degradation: A Potential Use of. *Planta medica* **61**, 510-514 (1995).

- 70 Bonadeo, I., Bottazzi, G. & Lavazza, M. Echinacin B: Active polysaccharide of the
Echinacea. *Riv. Ital. Essenza. Profumi. PianteOoff* **53**, 281-295 (1971).
- 71 Newall, C. A., Anderson, L. A. & Phillipson, J. D. *Herbal medicines. A guide for health-
care professionals*. (The pharmaceutical press, 1996).
- 72 Speroni, E., Govoni, P., Guizzardi, S., Renzulli, C. & Guerra, M. Anti-inflammatory and
cicatrizing activity of Echinacea pallida Nutt. root extract. *Journal of ethnopharmacology*
79, 265-272 (2002).
- 73 Tubaro, A., Tragni, E., Negro, P., Galli, C. & Loggia, R. D. Anti-inflammatory activity
of a polysaccharidic fraction of Echinacea angustifolia. *Journal of pharmacy and
pharmacology* **39**, 567-569 (1987).
- 74 Müller-Jakic, B., Breu, W., Pröbstle, A., Redl, K., Greger, H. & Bauer, R. In Vitro
Inhibition of Cyclooxygenase and 5-Lipoxygenase by Alkamides from Echinacea and
Achillea Species. *Planta Med* **60**, 37-40, doi:10.1055/s-2006-959404 (1994).
- 75 May, G. & Willuhn, G. Antiviral effect of aqueous plant extracts in tissue culture.
Arzneimittel-Forschung **28**, 1-7 (1977).
- 76 Cheminat, A., Zawatzky, R., Becker, H. & Brouillard, R. Caffeoyl conjugates from
Echinacea species: Structures and biological activity. *Phytochemistry* **27**, 2787-2794
(1988).
- 77 Beuscher, N., Bodinet, C., Willigmann, I. & Egert, D. Immune modulating properties of
root extracts of different Echinacea species. *Zeitschrift Phytotherapie* **16**, 157-166
(1995).
- 78 Thompson, K. D. Antiviral activity of Viracea® against acyclovir susceptible and
acyclovir resistant strains of herpes simplex virus. *Antiviral research* **39**, 55-61 (1998).
- 79 Wacker, A. & Hilbig, W. Virus-Inhibition by Echinacea purpurea. *Planta medica* **33**, 89-
102 (1978).
- 80 Dalby-Brown, L., Barsett, H., Landbo, A. R., Meyer, A. S. & Mølgaard, P. Synergistic
antioxidative effects of alkamides, caffeic acid derivatives, and polysaccharide fractions
from Echinacea purpurea on in vitro oxidation of human low-density lipoproteins.
Journal of agricultural and food chemistry **53**, 9413-9423 (2005).
- 81 Thygesen, L., Thulin, J., Mortensen, A., Skibsted, L. H. & Molgaard, P. Antioxidant
activity of cichoric acid and alkamides from Echinacea purpurea, alone and in
combination. *Food Chemistry* **101**, 74-81 (2007).
- 82 Hudec, J., Burdová, M., Kobida, L. U., Komora, L., Macho, V., Kogan, G., Turianica, I.,
Kochanová, R., Lozek, O. & Habán, M. Antioxidant capacity changes and phenolic
profile of Echinacea purpurea, nettle (*Urtica dioica* L.), and dandelion (*Taraxacum
officinale*) after application of polyamine and phenolic biosynthesis regulators. *Journal of
agricultural and food chemistry* **55**, 5689-5696 (2007).
- 83 Bauer, R. in *Immunomodulatory agents from plants*, Chemistry, analysis and
immunological investigations of Echinacea phytopharmaceuticals 41-88 (Springer,
1999).
- 84 Lucchesini, M., Bertoli, A., Mensuali-Sodi, A. & Pistelli, L. Establishment of in vitro
tissue cultures from Echinacea angustifolia DC adult plants for the production of
phytochemical compounds. *Scientia Horticulturae* **122**, 484-490 (2009).
- 85 Koroch, A., Juliani, H., Kapteyn, J. & Simon, J. In vitro regeneration of Echinacea
purpurea from leaf explants. *Plant cell, tissue and organ culture* **69**, 79-83 (2002).

- 86 Mustafa, N. R., De Winter, W., Van Iren, F. & Verpoorte, R. Initiation, growth and cryopreservation of plant cell suspension cultures. *Nature protocols* **6**, 715-742 (2011).
- 87 Morris, P. & Fowler, M. W. A new method for the production of fine plant cell suspension cultures. *Plant Cell, Tissue and Organ Culture* **1**, 15-24 (1981).
- 88 Guo, S., Man, S., Gao, W., Liu, H., Zhang, L. & Xiao, P. Production of flavonoids and polysaccharide by adding elicitor in different cellular cultivation processes of *Glycyrrhiza uralensis* Fisch. *Acta Physiologiae Plantarum* **35**, 679-686 (2013).
- 89 Jin, J. H., Shin, J. H., Kim, J. H., Chung, I. S. & Lee, H. J. Effect of chitosan elicitation and media components on the production of anthraquinone colorants in madder (*Rubia akane* Nakai) cell culture. *Biotechnology and Bioprocess Engineering* **4**, 300-304 (1999).
- 90 Wu, C., Tewari, R. K., Hahn, E. & Paek, K. Nitric oxide elicitation induces the accumulation of secondary metabolites and antioxidant defense in adventitious roots of *Echinacea purpurea*. *Journal of Plant Biology* **50**, 636-643 (2007).
- 91 Bota, C. & Deliu, C. The effect of copper sulphate on the production of flavonoids in *Digitalis lanata* cell cultures. *Farmacologia* **59**, 113-118 (2011).
- 92 Wu, C., Murthy, H. N., Hahn, E., Lee, H. L. & Paek, K. Efficient extraction of caffeic acid derivatives from adventitious roots of *Echinacea purpurea*. *Czech Journal of Food Sciences* **26**, 254-258 (2008).
- 93 Stanisavljević, I., Stojičević, S., Veličković, D., Veljković, V. & Lazić, M. Antioxidant and Antimicrobial Activities of *Echinacea* (*Echinacea purpurea* L.) Extracts Obtained by Classical and Ultrasound Extraction. *Chinese Journal of Chemical Engineering* **17**, 478-483 (2009).
- 94 Ainsworth, E. A. & Gillespie, K. M. Estimation of total phenolic content and other oxidation substrates in plant tissues using Folin–Ciocalteu reagent. *Nature protocols* **2**, 875-877 (2007).
- 95 Fang, N., Yu, S. & Prior, R. L. LC/MS/MS characterization of phenolic constituents in dried plums. *Journal of Agricultural and Food Chemistry* **50**, 3579-3585 (2002).
- 96 Ramachandra Rao, S. & Ravishankar, G. Plant cell cultures: chemical factories of secondary metabolites. *Biotechnology advances* **20**, 101-153 (2002).
- 97 Borenfreund, E. & Puerner, J. A. A simple quantitative procedure using monolayer cultures for cytotoxicity assays (HTD/NR-90). *Journal of tissue culture methods* **9**, 7-9 (1985).
- 98 Fotakis, G. & Timbrell, J. A. In vitro cytotoxicity assays: comparison of LDH, neutral red, MTT and protein assay in hepatoma cell lines following exposure to cadmium chloride. *Toxicology letters* **160**, 171-177 (2006).
- 99 Böyum, A. Isolation of mononuclear cells and granulocytes from human blood: Isolation of mononuclear cells by one centrifugation, and of granulocytes by combining centrifugation and sedimentation at 1 g. *Scandinavian journal of clinical and laboratory investigation. Supplementum* **97**, 77-89 (1967).
- 100 Instat, G. (1998).
- 101 Erel, O. A novel automated direct measurement method for total antioxidant capacity using a new generation, more stable ABTS radical cation. *Clinical Biochemistry* **37**, 277-285 (2004).
- 102 Athamna, A. & Ofek, I. Enzyme-linked immunosorbent assay for quantitation of attachment and ingestion stages of bacterial phagocytosis. *Journal of clinical microbiology* **26**, 62-66 (1988).

- 103 Simpson, D., Roth, R. & Loose, L. A rapid, inexpensive and easily quantified assay for phagocytosis and microbicidal activity of macrophages and neutrophils. *Journal of immunological methods* **29**, 221-226 (1979).
- 104 Nauseef, W. M. in *Neutrophil Methods and Protocols*, Isolation of human neutrophils from venous blood 15-20 (Springer, 2007).
- 105 Manosroi, A., Saraphanchotiwiththaya, A. & Manosroi, J. In vitro immunomodulatory effect of *Pouteria cambodiana* (Pierre ex Dubard) Baehni extract. *Journal of ethnopharmacology* **101**, 90-94 (2005).
- 106 Froebel, K., Pakker, N., Aiuti, F., Bofill, M., Choremi-Papadopoulou, H., Economidou, J., Rabian, C., Roos, M., Ryder, L. & Miedema, F. Standardisation and quality assurance of lymphocyte proliferation assays for use in the assessment of immune function. *Journal of immunological methods* **227**, 85-97 (1999).
- 107 Boscolo, P., Del Signore, A., Sabbioni, E., Di Gioacchino, M., Di Giampaolo, L., Reale, M., Conti, P., Paganelli, R. & Giaccio, M. Effects of resveratrol on lymphocyte proliferation and cytokine release. *Annals of Clinical & Laboratory Science* **33**, 226-231 (2003).
- 108 Delgado, I. & Paumgarten, F. Effects of *Euphorbia milii* latex on mitogen-induced lymphocyte proliferation. *Revista Brasileira de Plantas Medicinai* **16**, 107-111 (2014).
- 109 Hendra, R., Ahmad, S., Oskoueian, E., Sukari, A. & Shukor, M. Y. Antioxidant, anti-inflammatory and cytotoxicity of *Phaleria macrocarpa* (Boerl.) Scheff Fruit. *BMC complementary and alternative medicine* **11**, 110 (2011).
- 110 Ding, A. H., Nathan, C. F. & Stuehr, D. J. Release of reactive nitrogen intermediates and reactive oxygen intermediates from mouse peritoneal macrophages. Comparison of activating cytokines and evidence for independent production. *The Journal of Immunology* **141**, 2407-2412 (1988).
- 111 Pellati, F., Benvenuti, S., Magro, L., Melegari, M. & Soragni, F. Analysis of phenolic compounds and radical scavenging activity of *Echinacea* spp. *Journal of Pharmaceutical and Biomedical Analysis* **35**, 289-301 (2004).
- 112 Rininger, J. A., Kickner, S., Chigurupati, P., Mclean, A. & Franck, Z. Immunopharmacological activity of *Echinacea* preparations following simulated digestion on murine macrophages and human peripheral blood mononuclear cells. *Journal of Leukocyte Biology* **68**, 503-510 (2000).
- 113 Zhai, Z., Haney, D., Wu, L., Solco, A., Murphy, P. A., Wurtele, E. S., Kohut, M. L. & Cunnick, J. E. Alcohol extracts of *Echinacea* inhibit production of nitric oxide and tumor necrosis factor- α by macrophages in vitro. *Food and agricultural immunology* **18**, 221-236 (2007).
- 114 Chaves, F., Chacón, M., Badilla, B. & Arévalo, C. Effect of *Echinacea purpurea* (Asteraceae) aqueous extract on antibody response to *Bothrops asper* venom and immune cell response. *Revista de biología tropical* **55**, 113-119 (2007).
- 115 Siddiqui, Z. H., Mujib, A., Aslam, J., Hakeem, K. R. & Parween, T. in *Crop Improvement: New Approaches and Modern Techniques*, In vitro Production of Secondary Metabolites Using Elicitor in *Catharanthus roseus*: A Case Study 401-419 (Springer, 2013).
- 116 Li, W. & Barz, W. Biotechnological production of two new 8, 4'-oxynorneolignans by elicitation of *Echinacea purpurea* cell cultures. *Tetrahedron letters* **46**, 2973-2977 (2005).

دراسة بالتقنية الحيوية لنبات الاشنيسيا المنزرع محليا

رسالة مقدمة إلى

كلية الصيدلة- جامعة الإسكندرية

كمطلب جزئى للحصول على

درجة الماجستير في العلوم الصيدلانية (عقاقير)

مقدمة من

نيقين ممدوح عبد الرحمن على

بكالوريوس فى العلوم الصيدلانية

كلية الصيدلة

جامعة الإسكندرية

٢٠٠٨

قسم العقاقير

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٢٠١٤

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و القائم بأعمال رئيس مركز تطوير الصناعات الدوائية و الصيدلية و التخمرية
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للحصول على درجة
الماجستير في العلوم الصيدلانية
(عقاقير)

موافق

لجنة المناقشة و الحكم على الرسالة

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